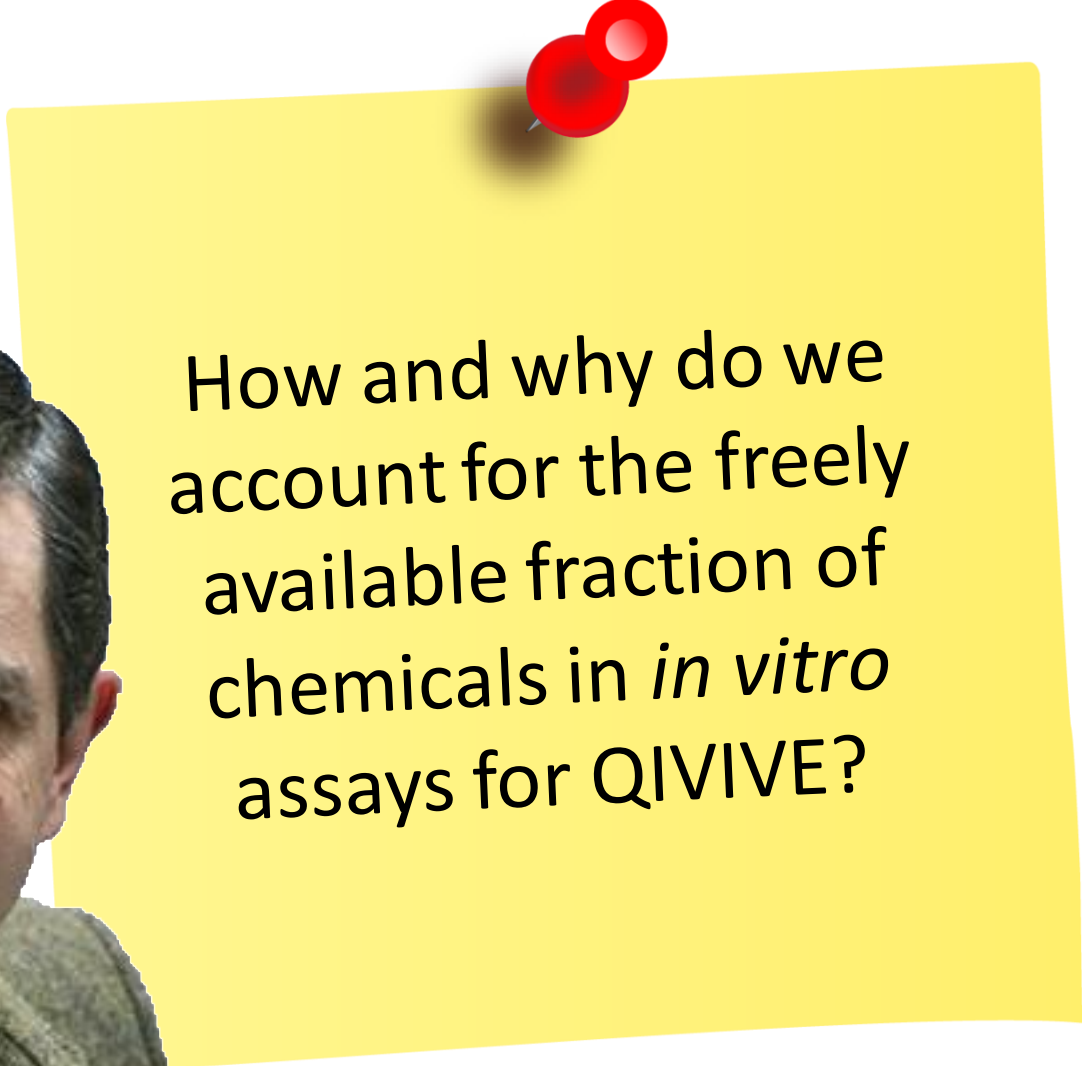


# Measuring and modeling the distribution of test chemicals in in vitro toxicity assays

Nynke Kramer<sup>1</sup>

1. Toxicology Division, Wageningen University, Wageningen, The Netherlands

# Presentation Aim

A yellow sticky note is pinned to the slide with two red pushpins. The note contains the text 'How and why do we account for the freely available fraction of chemicals in *in vitro* assays for QIVIVE?'.

How and why do we account for the freely available fraction of chemicals in *in vitro* assays for QIVIVE?





# Basis of Toxicity Testing in the 21<sup>st</sup> Century: *In Vitro* Cell Assays



Mechanistic approach



Human tissue/tissue of species of interest



High throughput



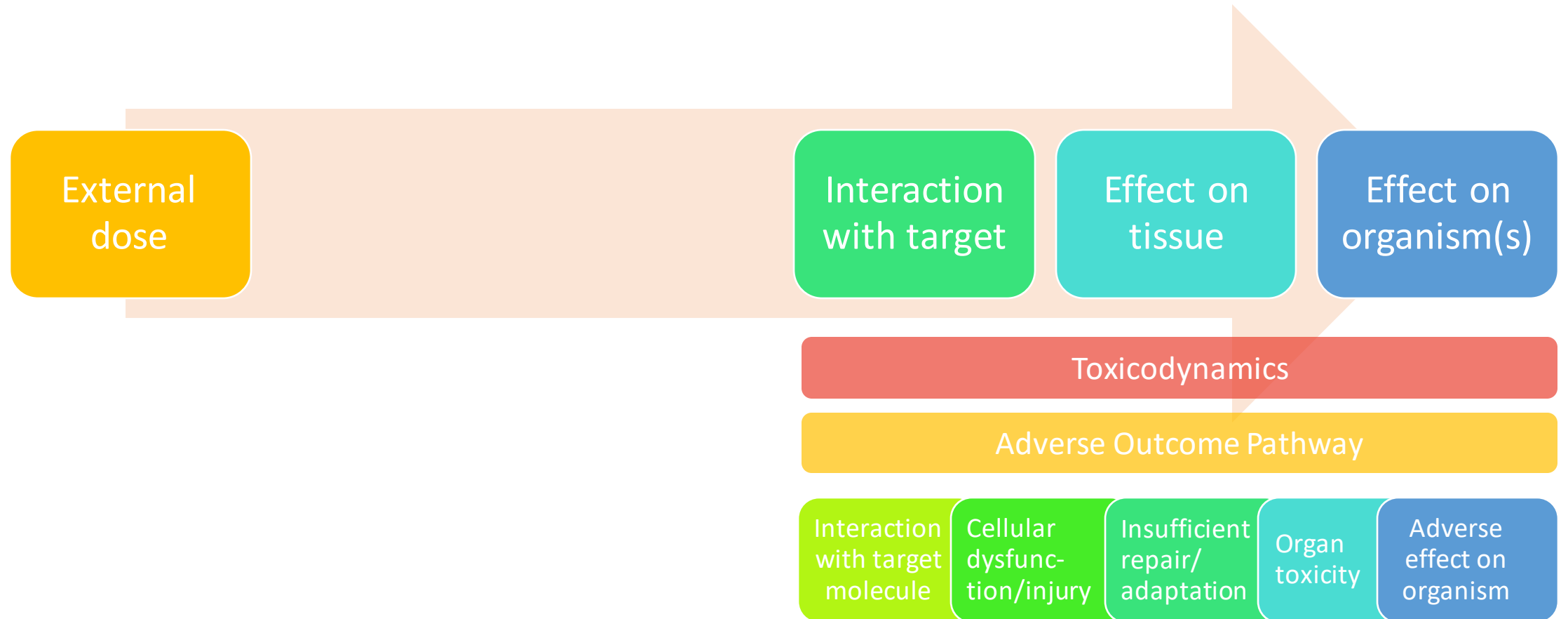
Little waste



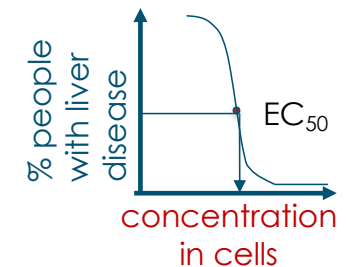
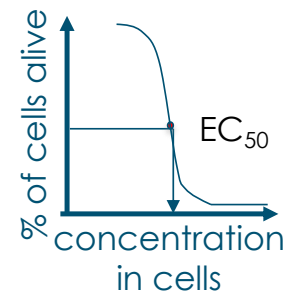
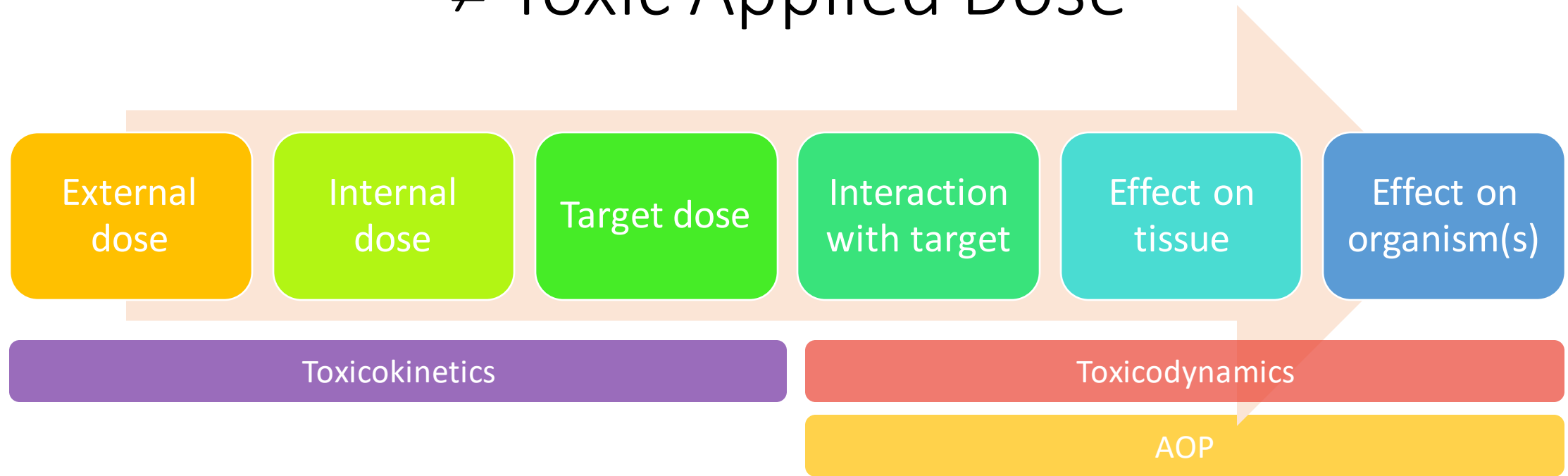
(Ethically) sound science

3Rs: replacement,  
reduction, refinement

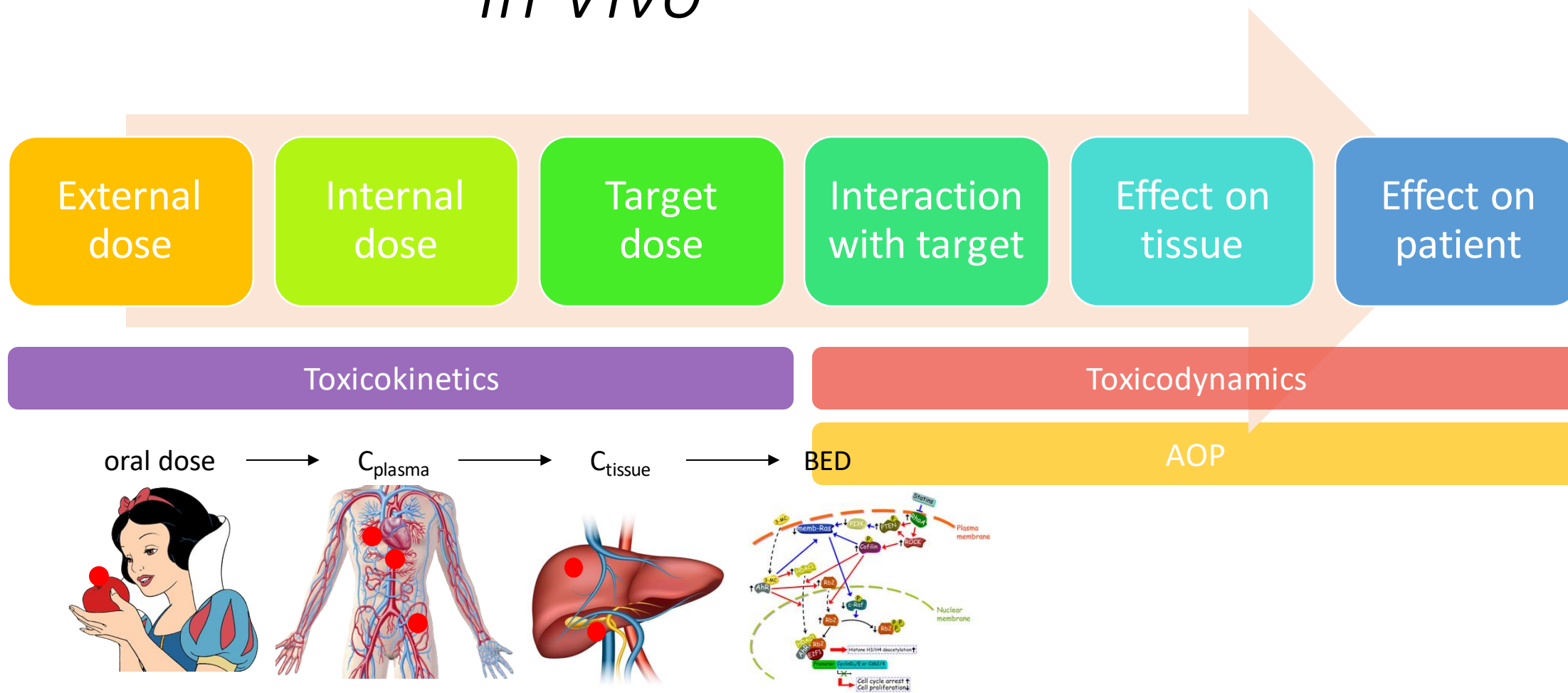
# *In Vitro* Assays in Toxicity Testing in the 21<sup>st</sup> Century



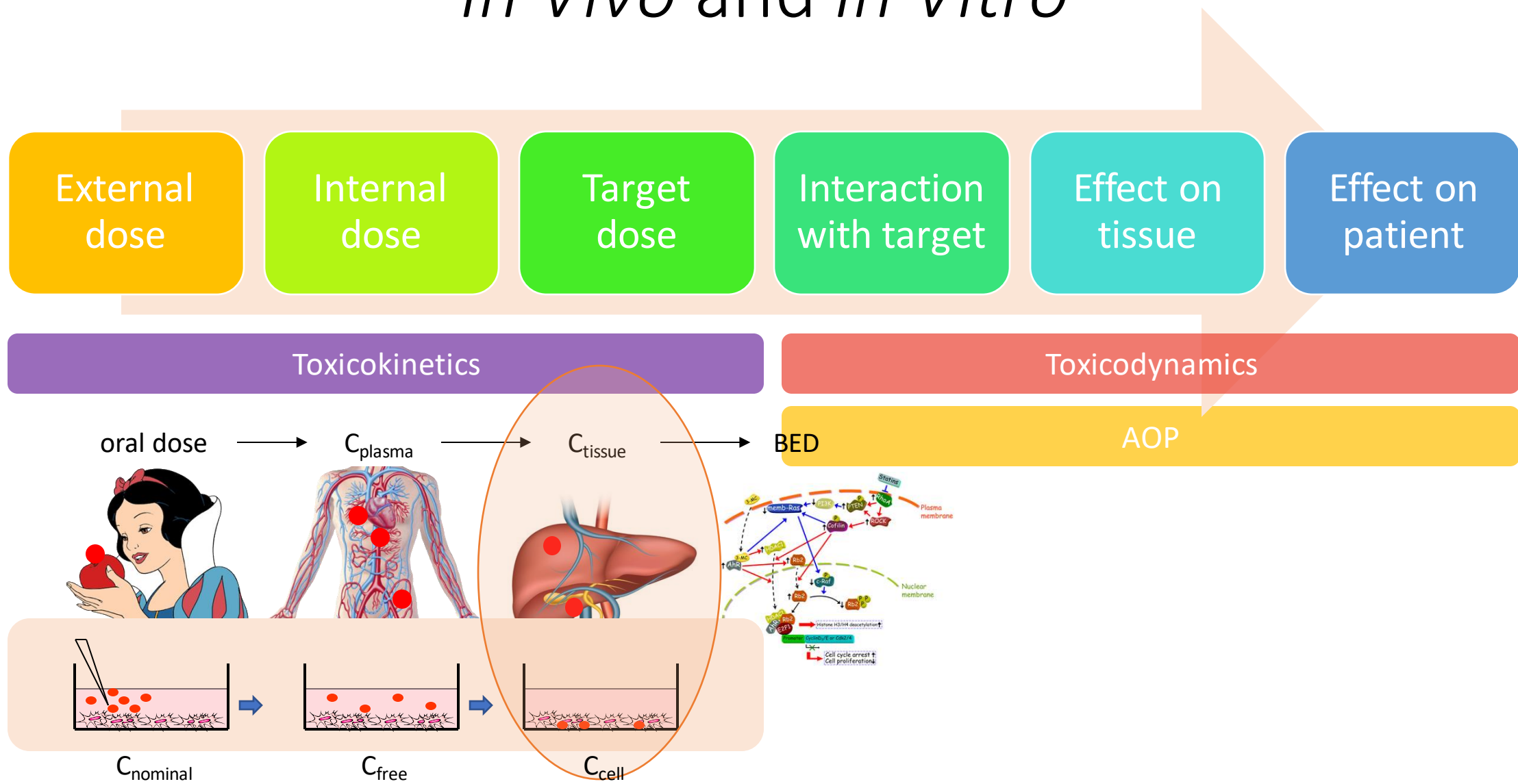
# Toxic Concentration *In Vitro* ≠ Toxic Applied Dose



# Need to Account for Kinetics *In Vivo*



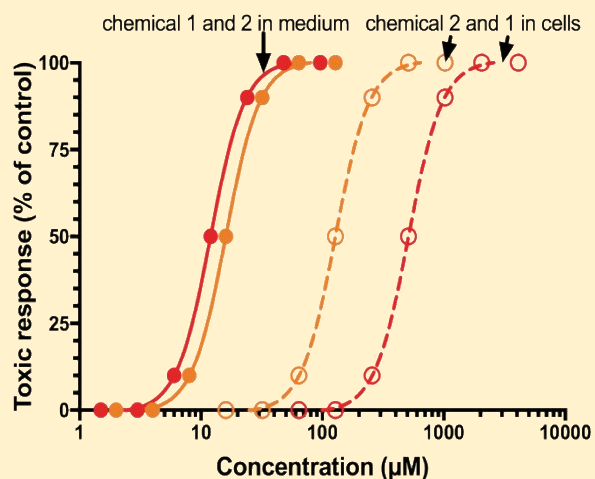
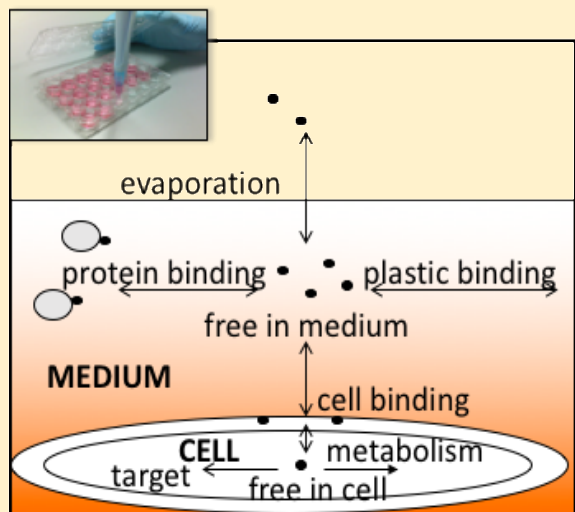
# Need to Account for Kinetics *In Vivo* and *In Vitro*





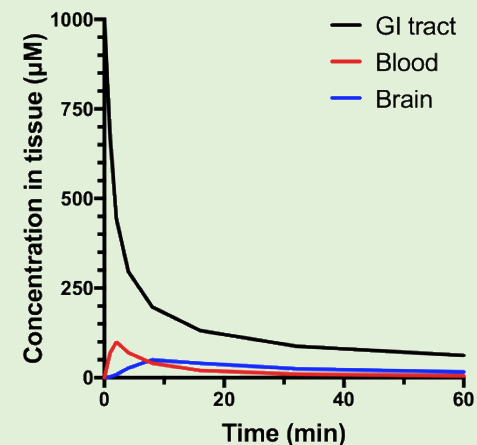
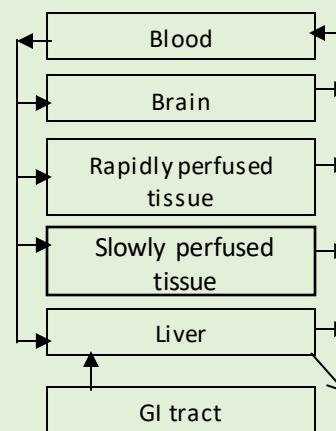
# Tools for Accounting for Kinetics

## *In vitro* kinetics



*In Vitro* Distribution Kinetics Models

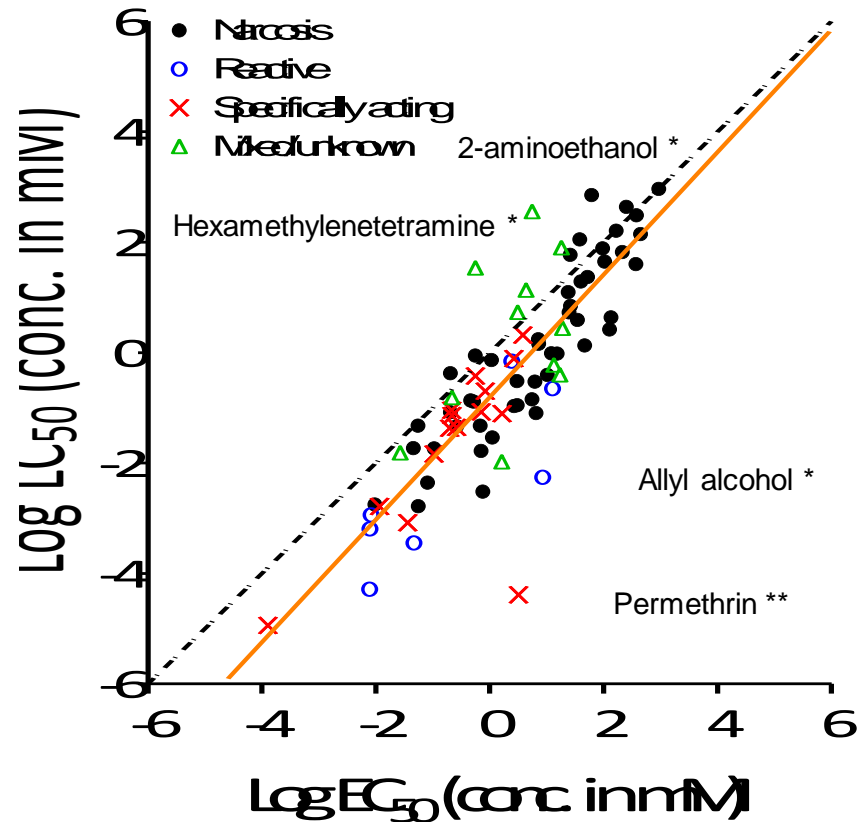
## *In vivo* kinetics



Physiologically Based Kinetic Models



# More simple forms of QIVIVE illustrate importance of understanding *in vitro* kinetics

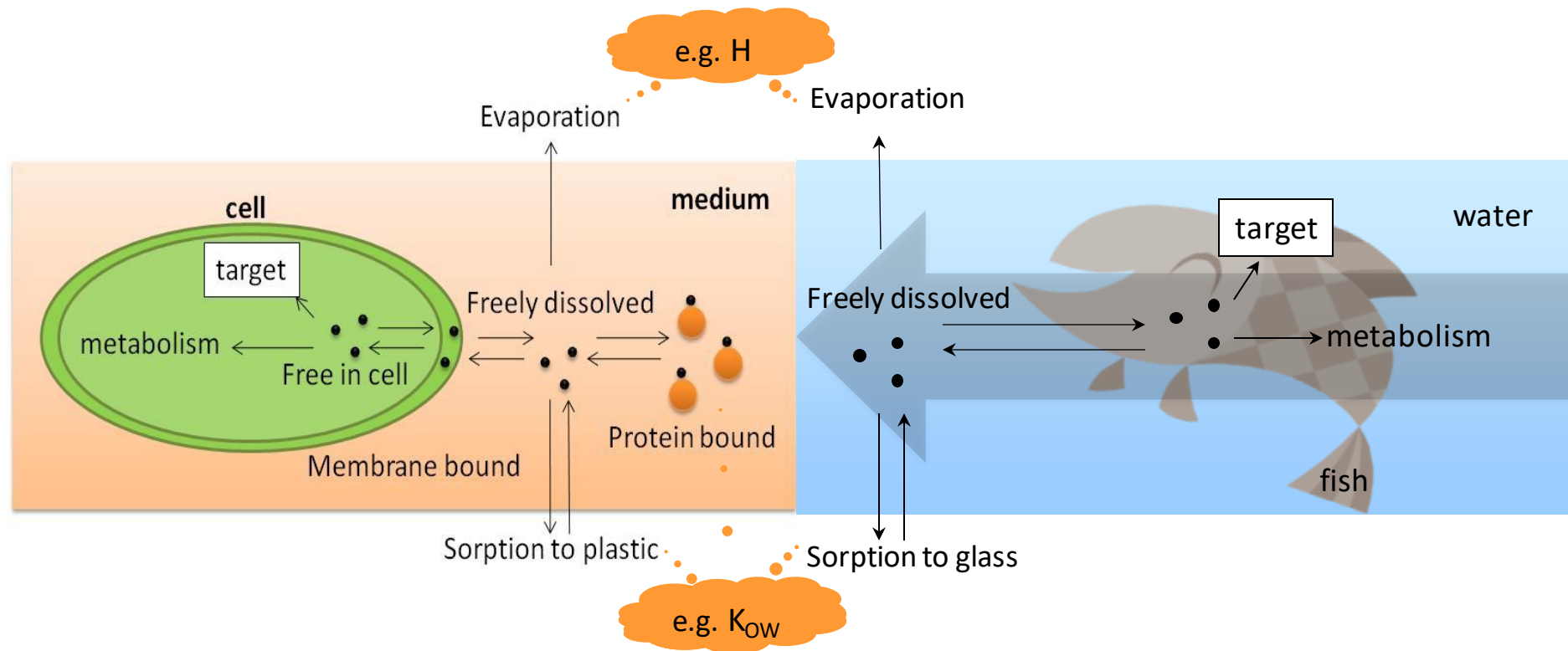


- *In vitro* basal cytotoxicity assay (Halle Database) vs. acute fish bioassay (EPA Duluth FHM Database)
- Good correlation, but high variability and poor sensitivity
- Outliers specifically acting, bioactivated chemicals
- Correlation improves when only narcotic chemicals are used in regression
- *In vitro* still less sensitive than fish acute bioassay

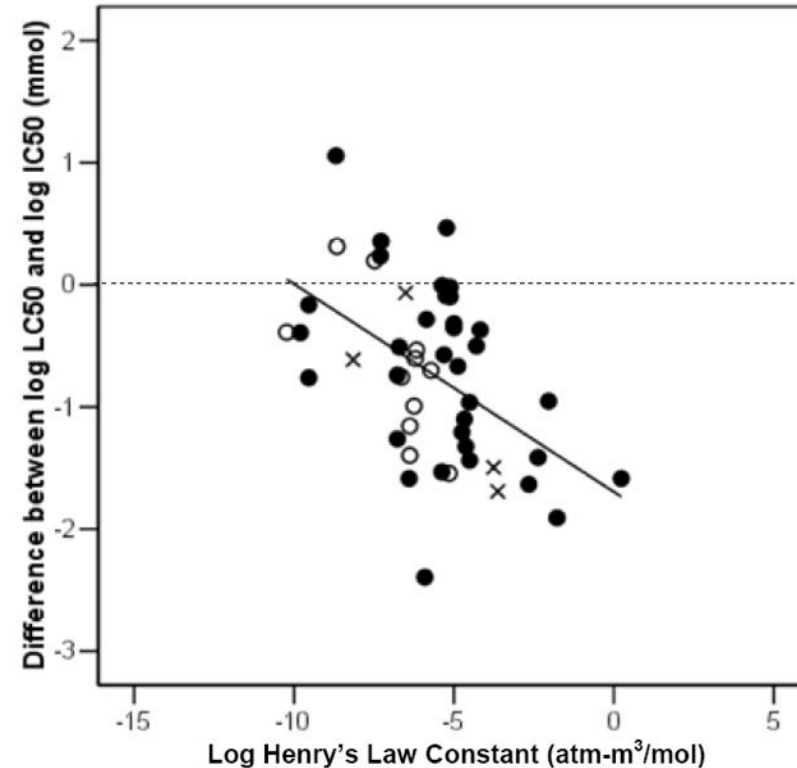
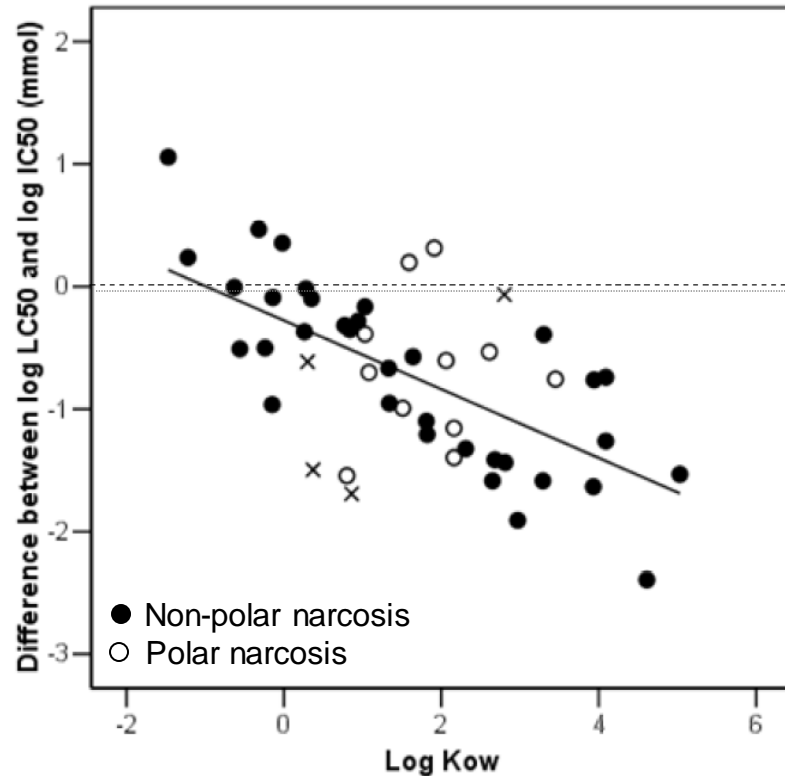
$$\text{Log LC}_{50} (\text{mM}) = 1.10 \pm 0.08 \text{ Log IC}_{50} (\text{mM}) - 0.81 \pm 0.11, R^2 = 0.70.$$

$$\text{Log LC}_{50} (\text{mM}) = 1.11 \pm 0.08 \text{ Log IC}_{50} (\text{mM}) - 0.81 \pm 0.12, R^2 = 0.80.$$

# Importance of *In Vitro* Kinetics in QIVIVE



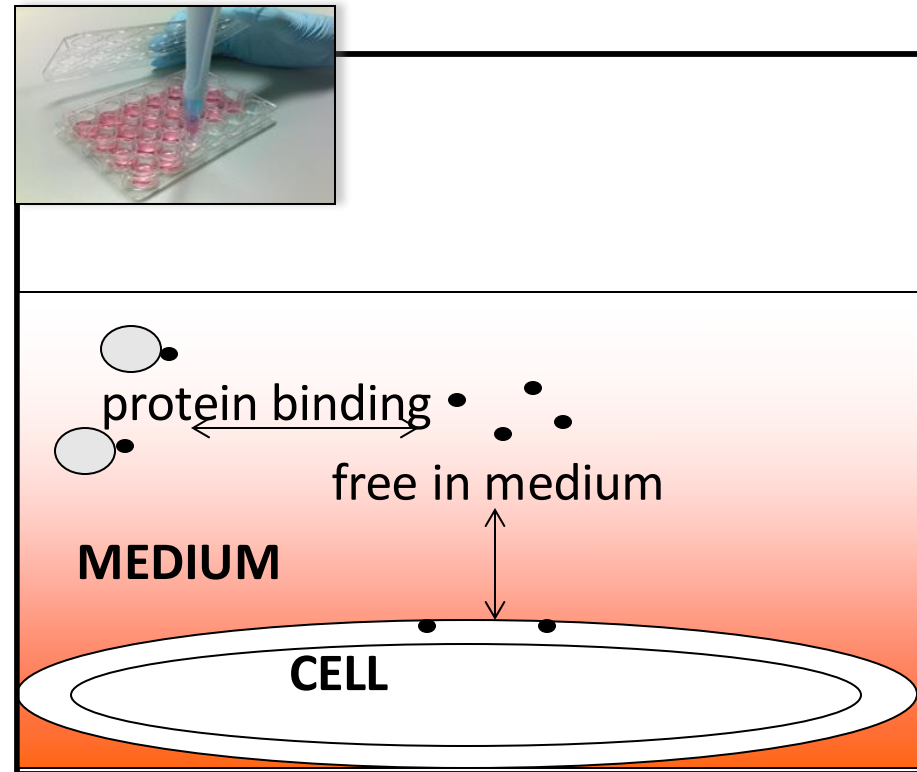
# Role of Physicochemical Properties



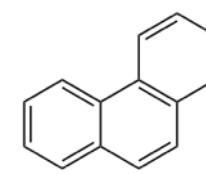
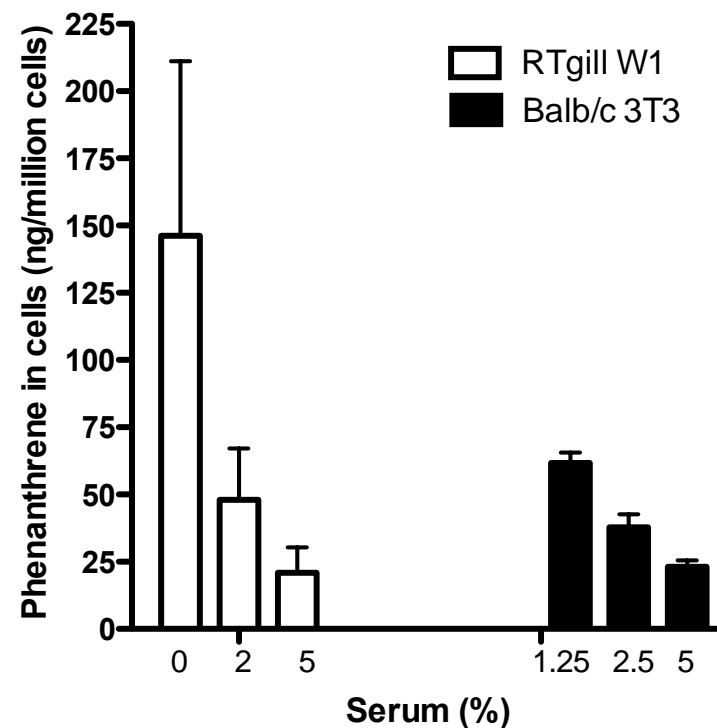
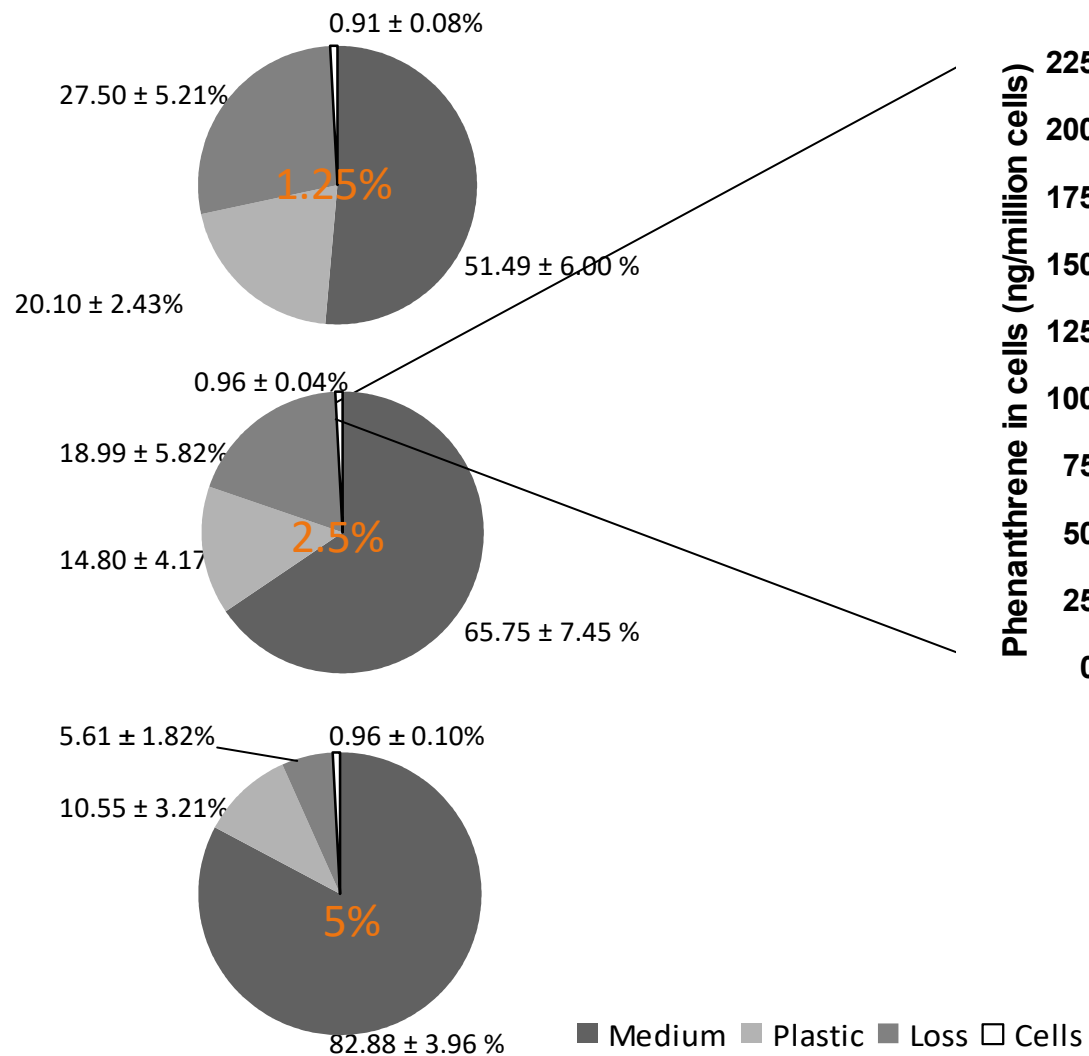
$$\text{Log LC50} - \text{Log IC50 (mM)} = -0.26 (\pm 0.05) \text{ Log } K_{ow} - 0.30 (\pm 0.11), R^2 = 0.38$$

$$\text{Log LC50} - \text{Log IC50 (mM)} = -0.17 (\pm 0.04) \text{ Log } H (\text{atm-m}^3/\text{mol}) - 1.70 (\pm 0.26), R^2 = 0.25$$

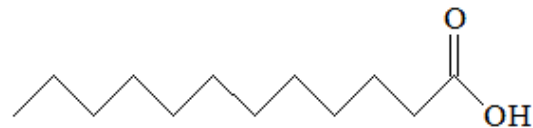
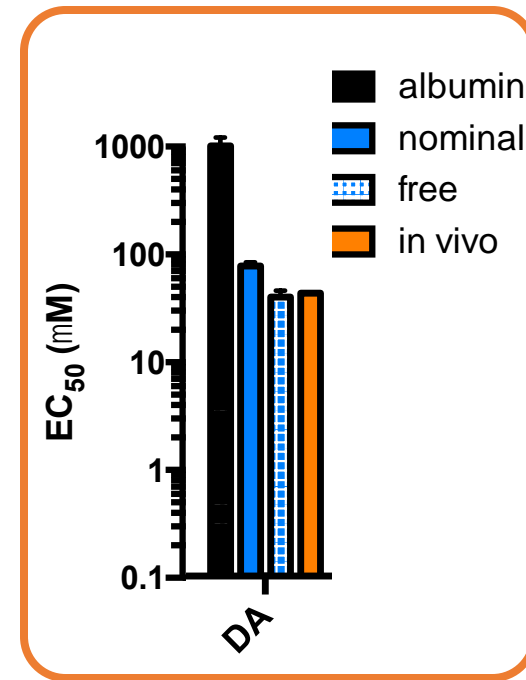
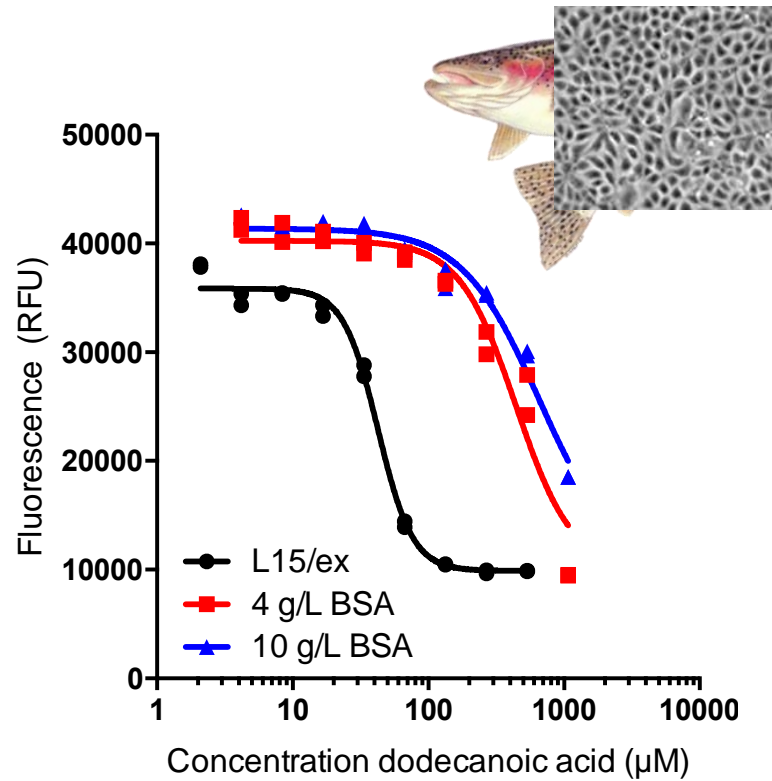
# Determinants of *In Vitro* Distribution Kinetics: Serum Constituent Binding



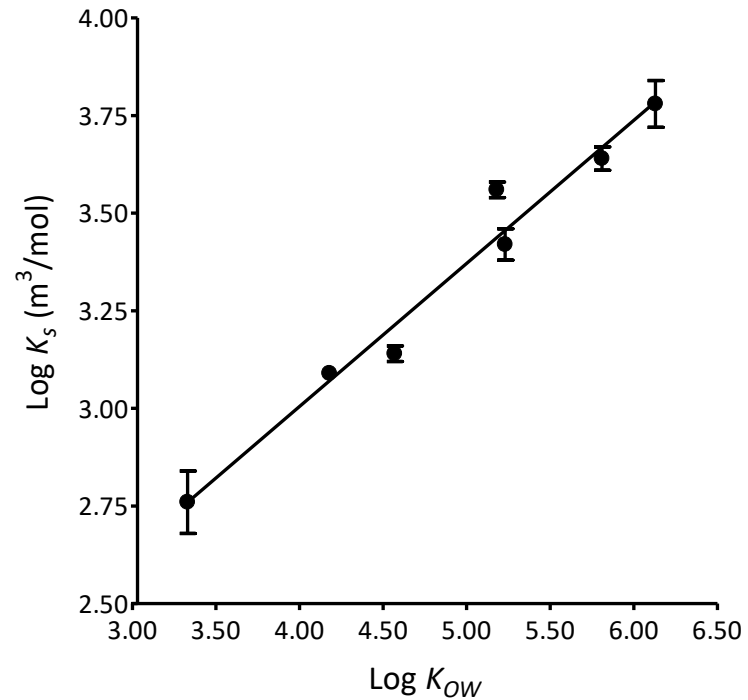
# Serum Constituent Binding



# Serum Constituent Binding



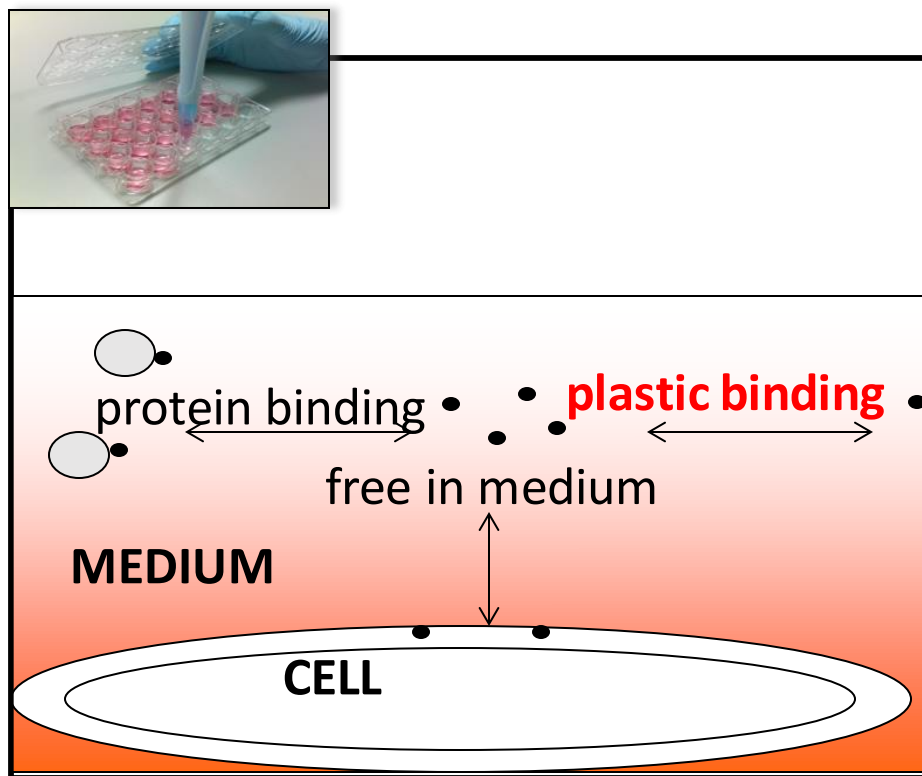
# Serum Constituent Binding



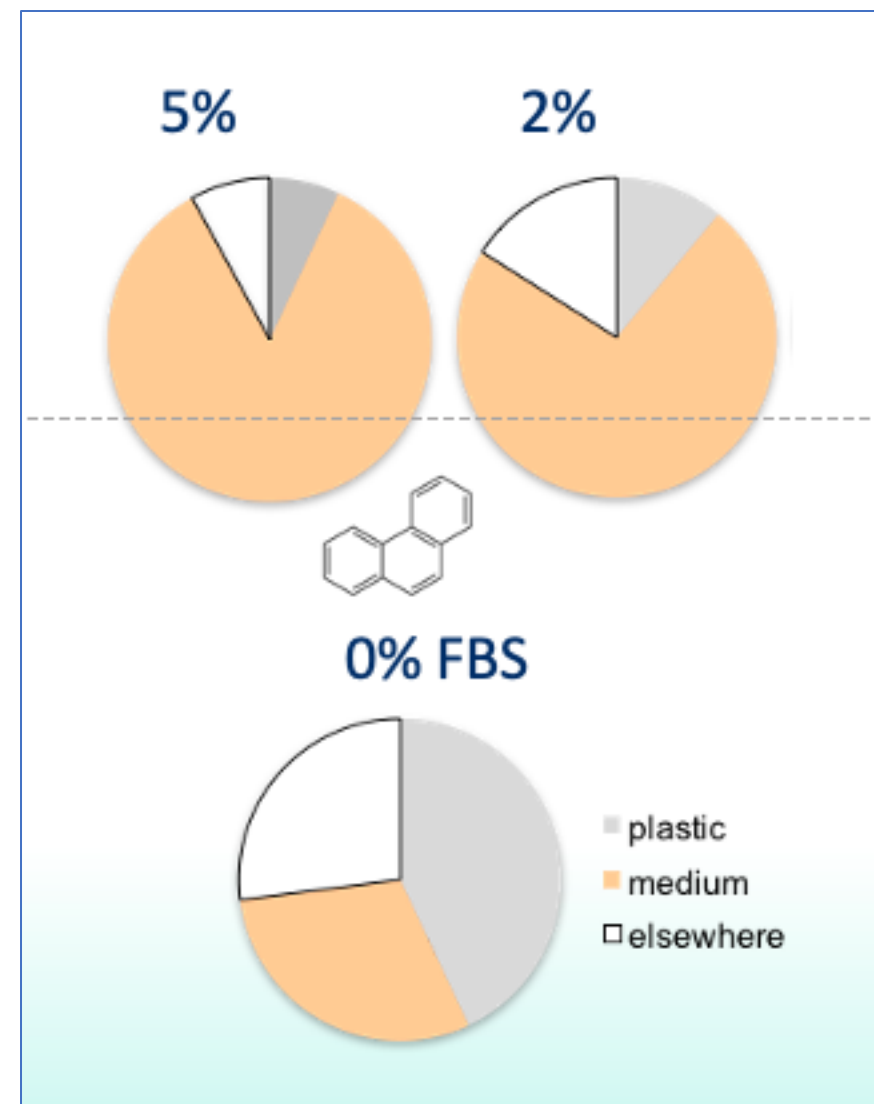
Quantitative Structure Property Relationships (QSPR)



# Well Plate Plastic Binding

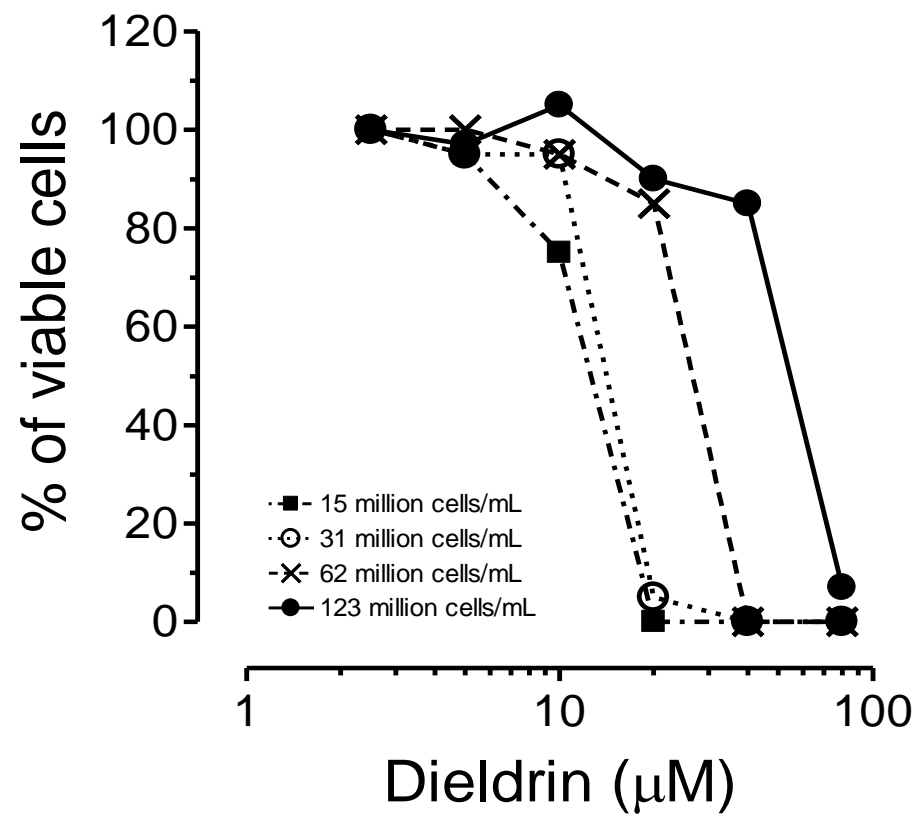
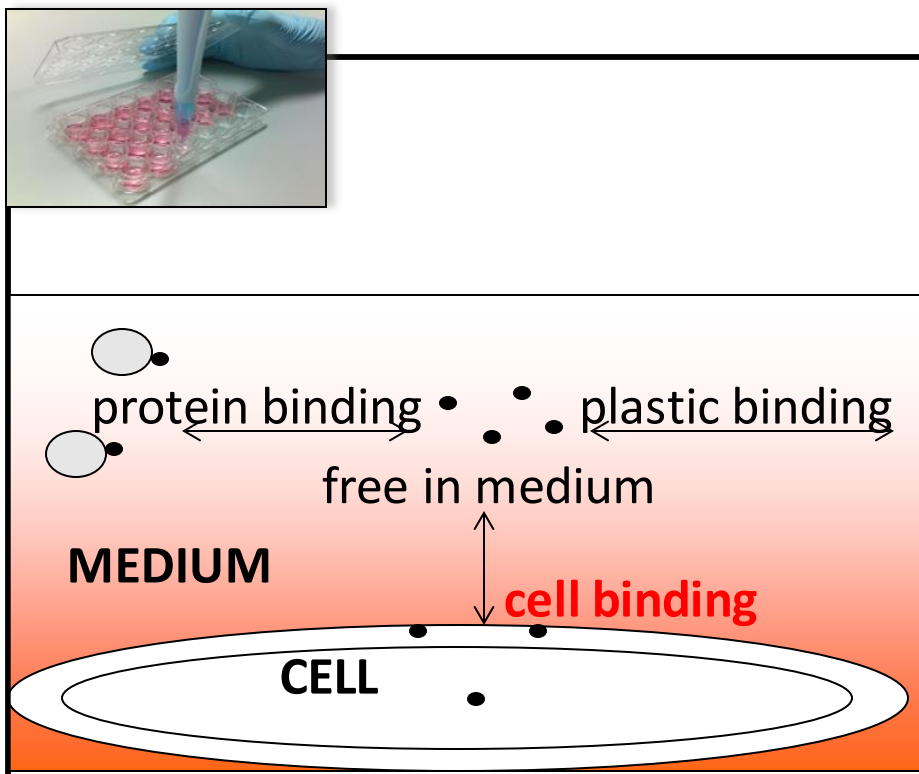


Kramer et al. (2012) Chem. Res. Toxicol. 25, 436  
 Schaap et al., manuscript in preparation

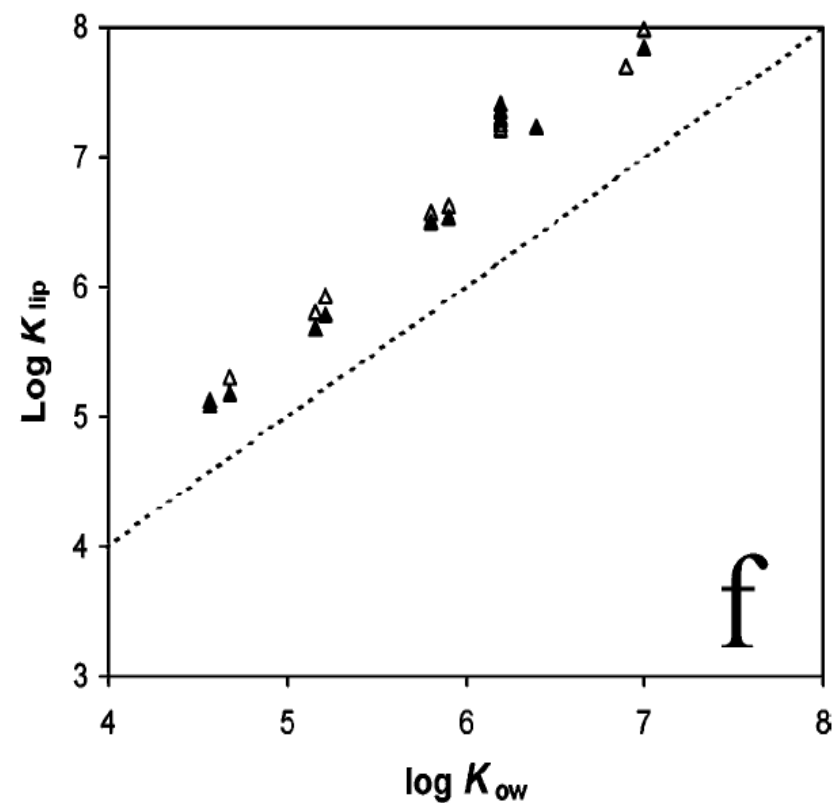
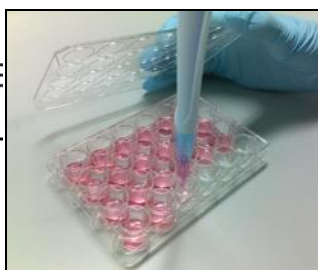
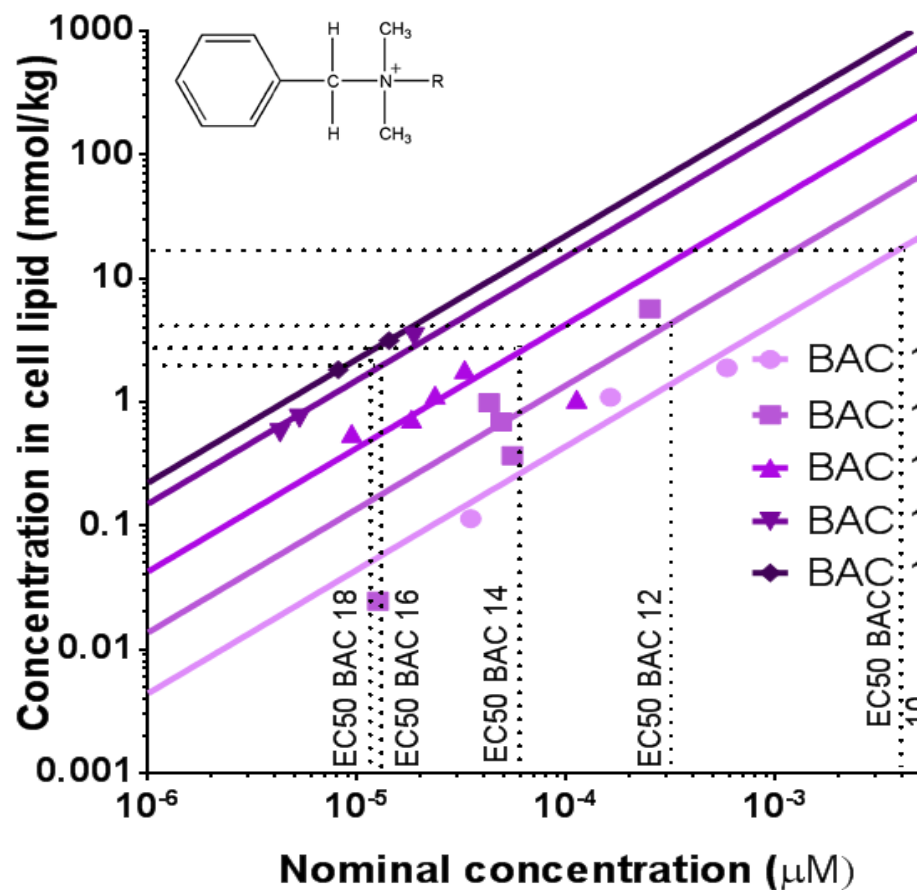
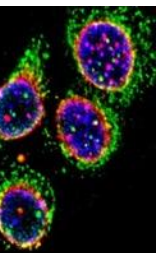




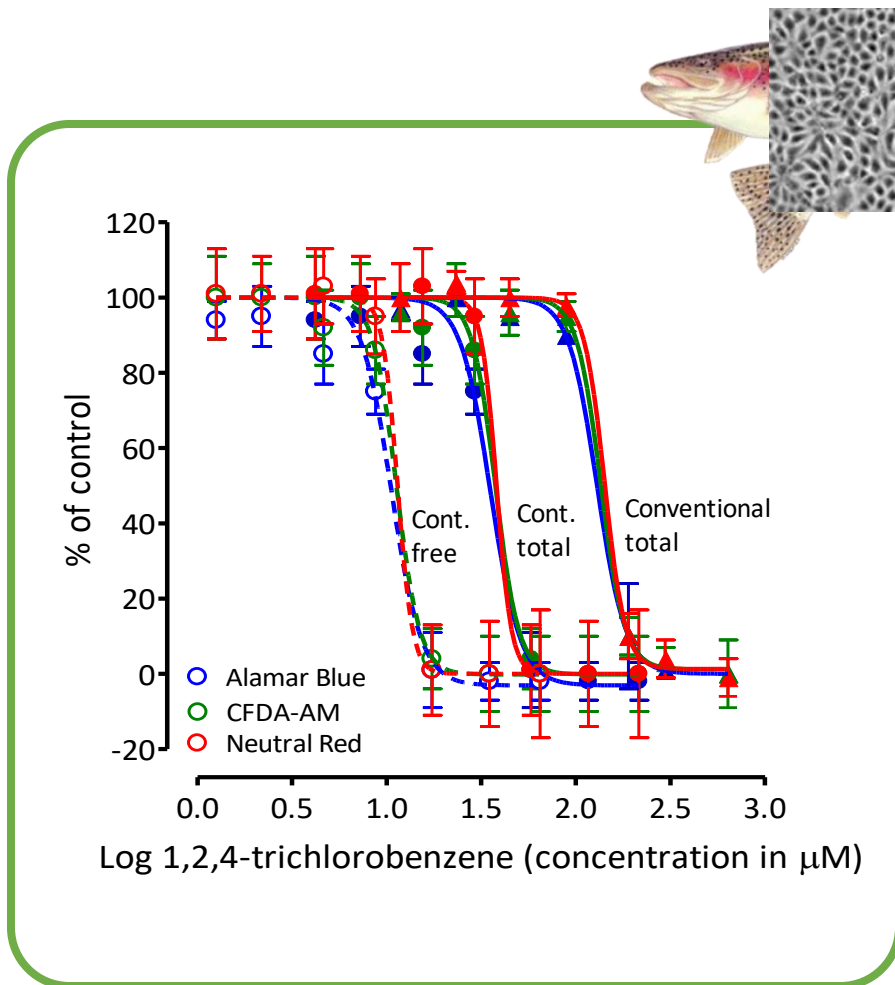
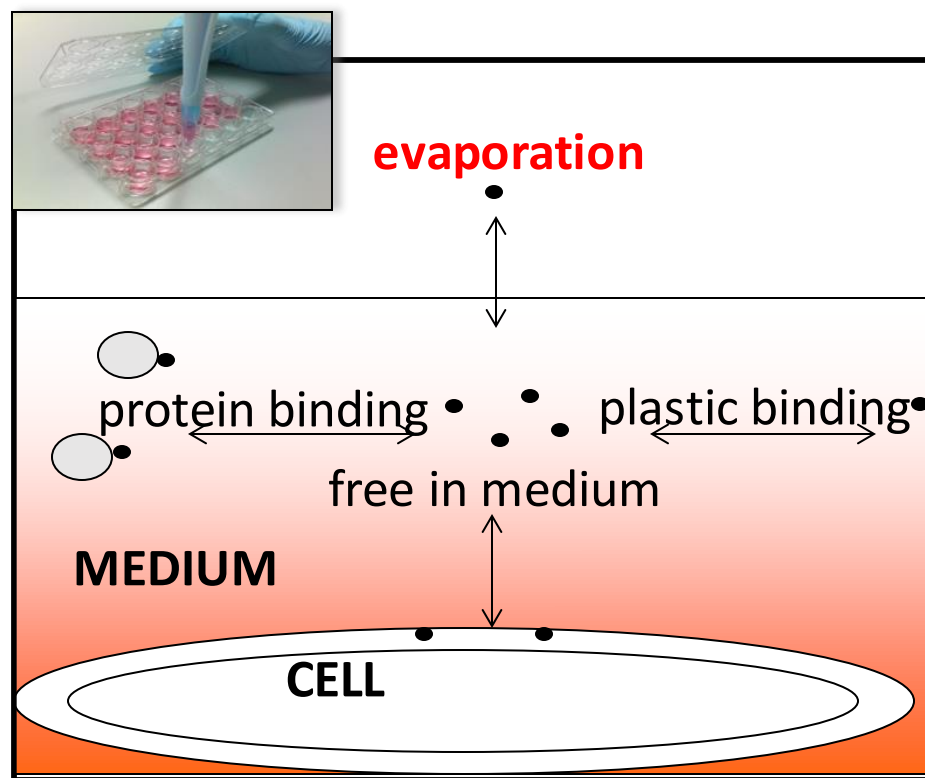
# Cell Association



# Cell Binding Affinity vs. QIVIVE

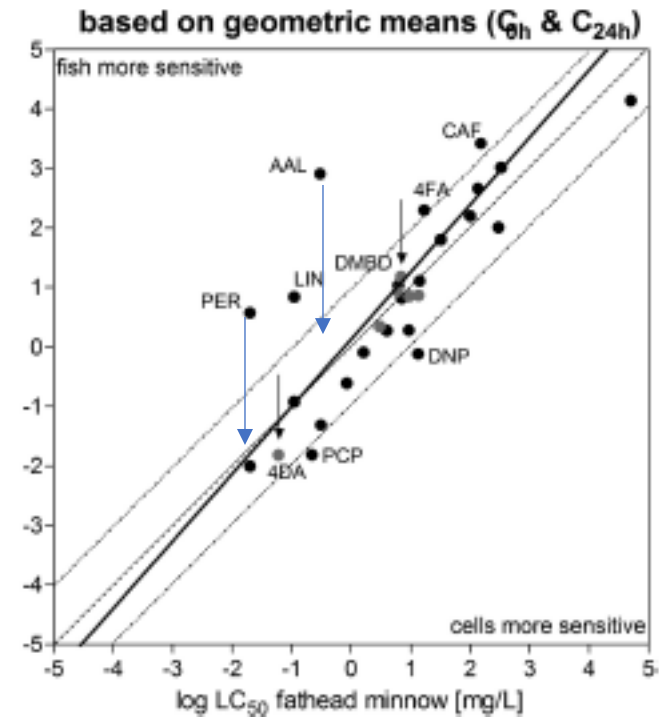
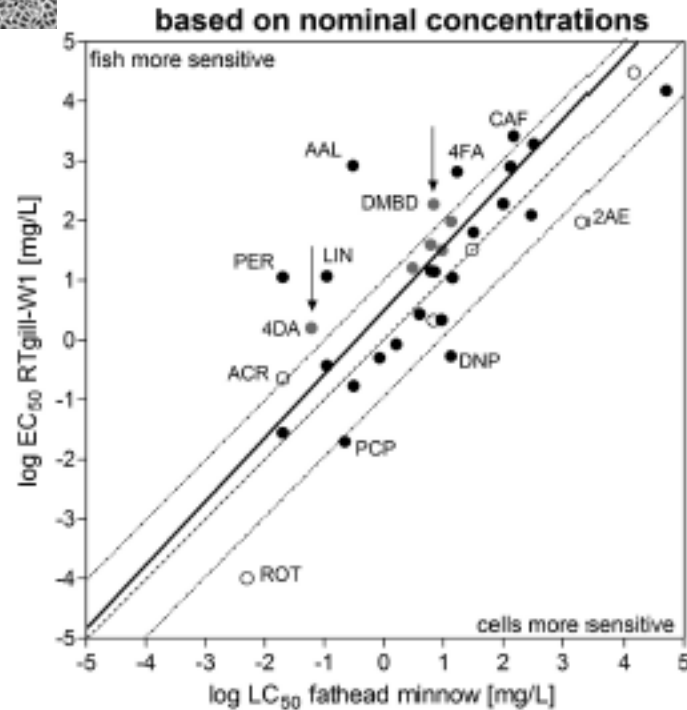
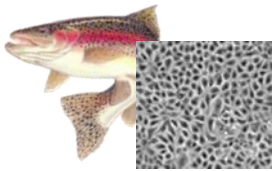


# Evaporation



Recovery from medium after 48h	Conventional dosing: 11% Continuous dosing: 105%
EC <sub>50</sub> ( $\mu\text{M}$ )	Conventional: 135 Continuous total: 38 Continuous free: 11 Fathead Minnow: 16
Bound to Serum Constituents	70%

# Importance of *In Vitro* Distribution Kinetics in QIVIVE

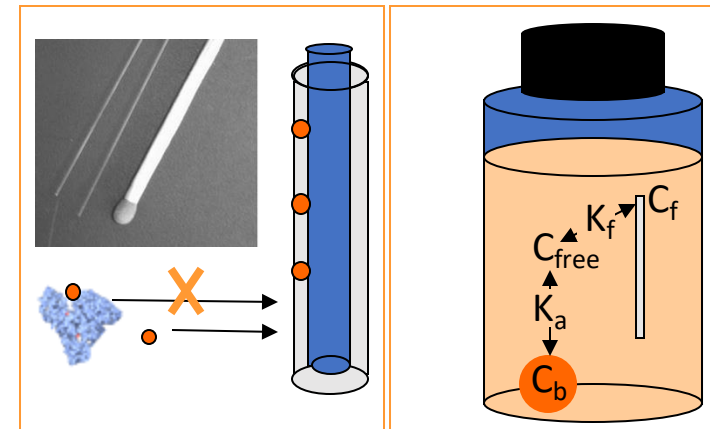


Halle 2003

Basker & Murthy 2018

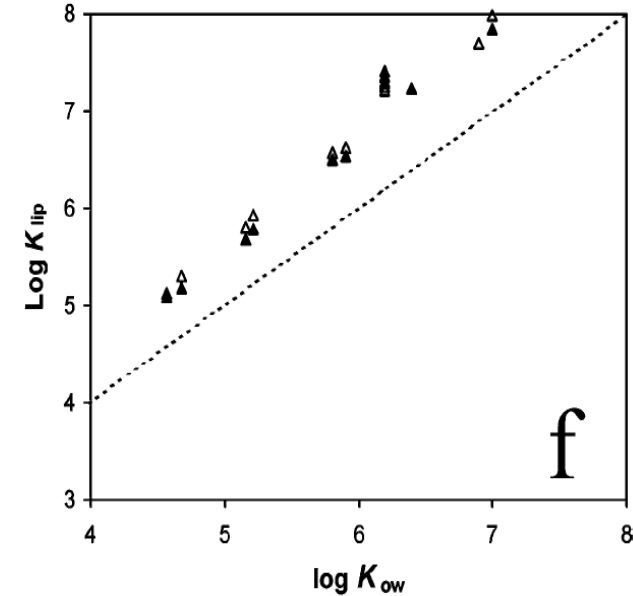
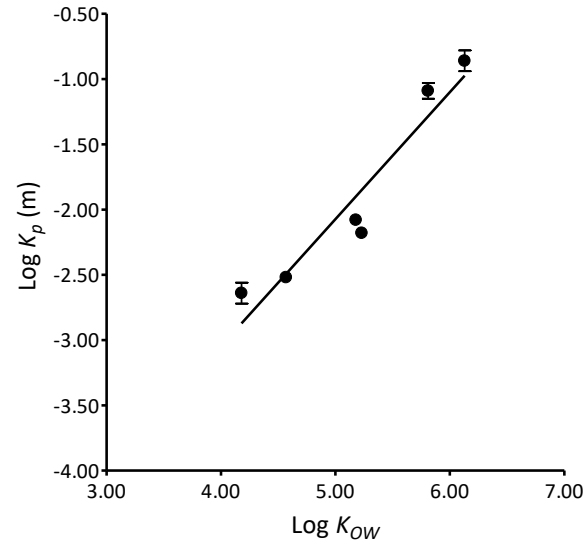
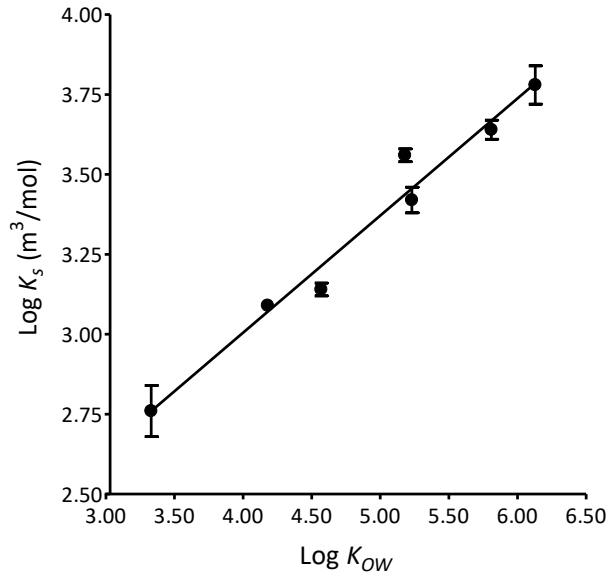
# Analytical Methods for Assessing *In Vitro* Distribution Kinetics

- Rapid Equilibrium Dialysis (RED)
- Ultrafiltration
- Ultracentrifugation
- Column Chromatography
- Solid Phase (Micro)extraction (SPME)
- Methods discussed in Groothuis et al. (2015) *Toxicology In Vitro* 332, 30-40.





# Modelling *In Vitro* Distribution Kinetics

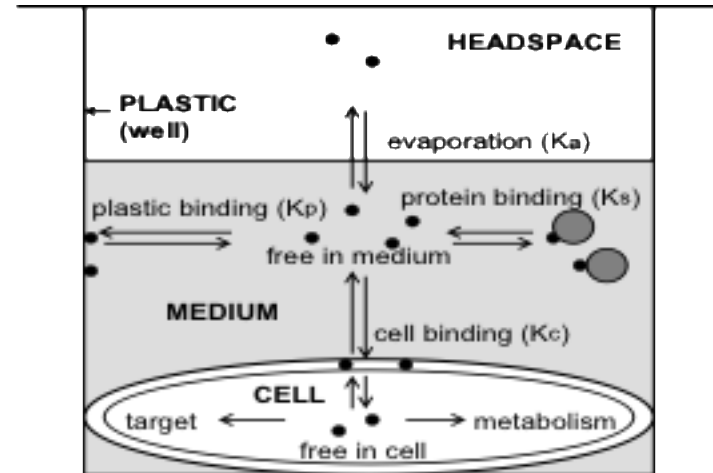


$$F = \frac{1}{1 + K_s \cdot [S] + K_p \cdot [P] + K_c \cdot [C] + K_a \cdot \frac{V_a}{V_m}}$$

Schaap et al. (in preparation).

Jonker et al (2007) Environ. Sci. Technol. 41, 7363

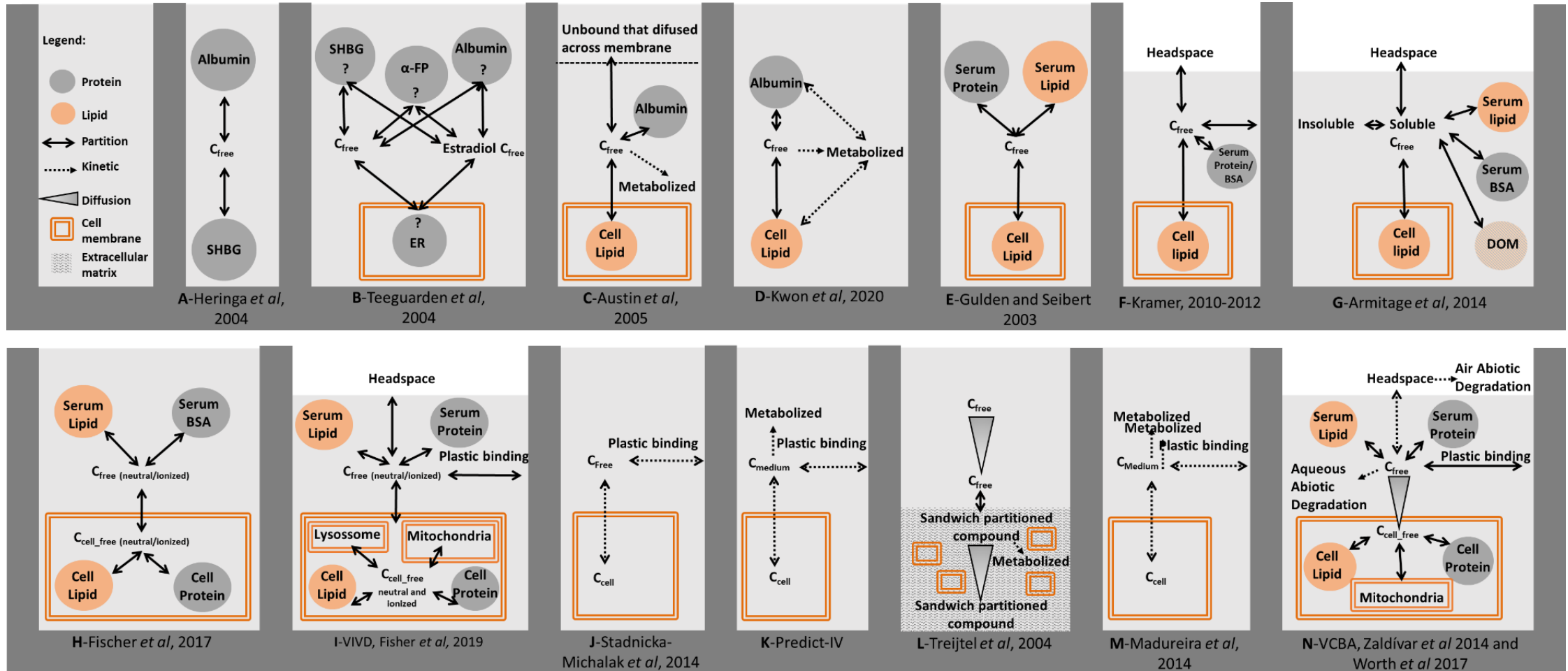
# Modelling *In Vitro* Distribution Kinetics



$$F = \frac{1}{1 + K_s [S] + K_p [P] + K_c [C] + K_a [A]}$$

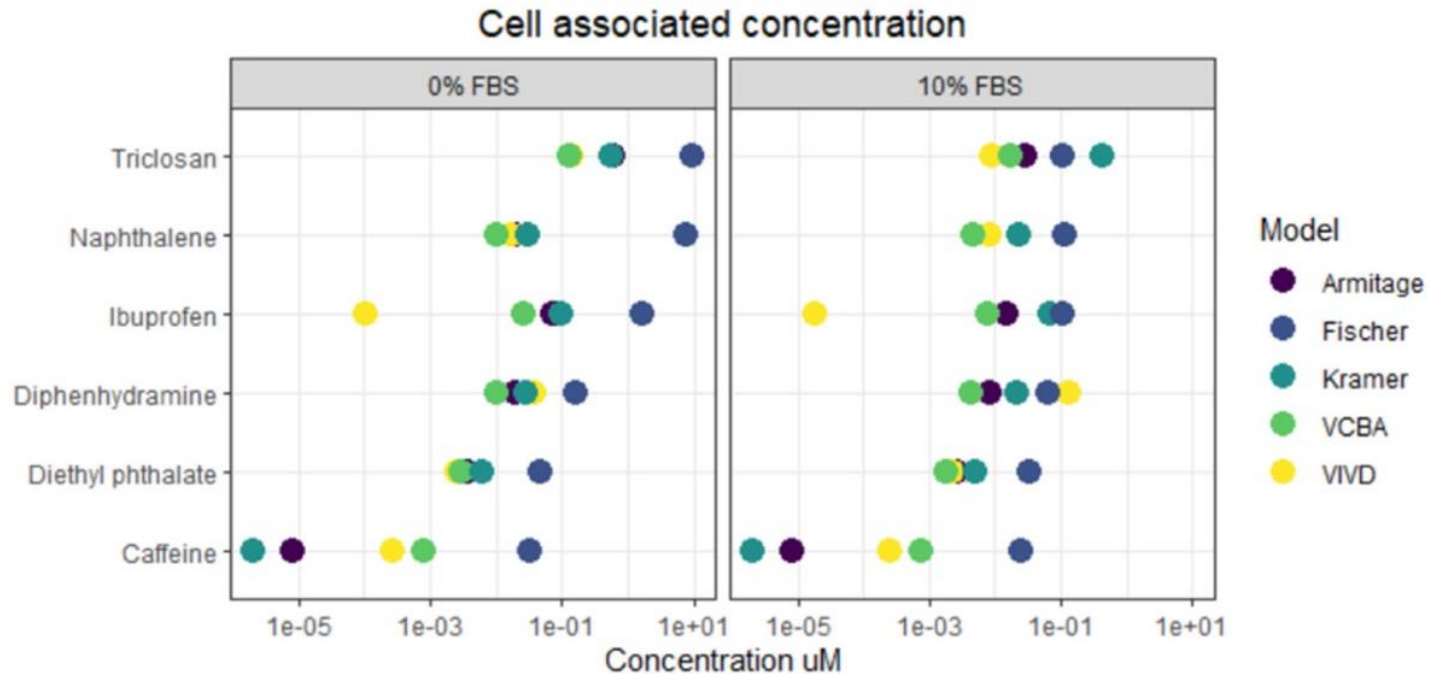
$$F = \frac{1}{1 + 10^{0.37 \log K_{ow} - 0.29} [S] + 10^{0.97 \log K_{ow} - 6.94} [P] + 10^{1.25 \log K_{ow} - 3.70} [C] + \frac{H}{8.3144T} \cdot \frac{V_a}{V_m}}$$

# Modeling *In Vitro* Distribution Kinetics



# Comparing Distribution Predictions

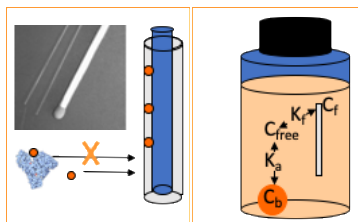
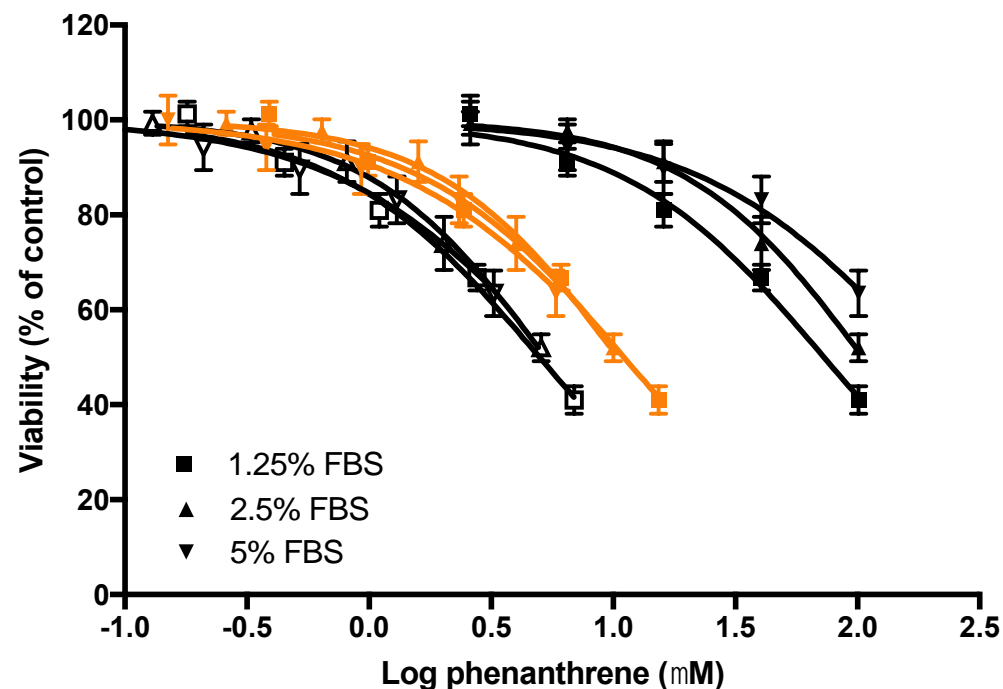
Simulating distribution kinetics of test chemicals at 100  $\mu\text{M}$  nominal concentrations in *in vitro* assays with HepaRG (7,600 cell/cm<sup>2</sup>)



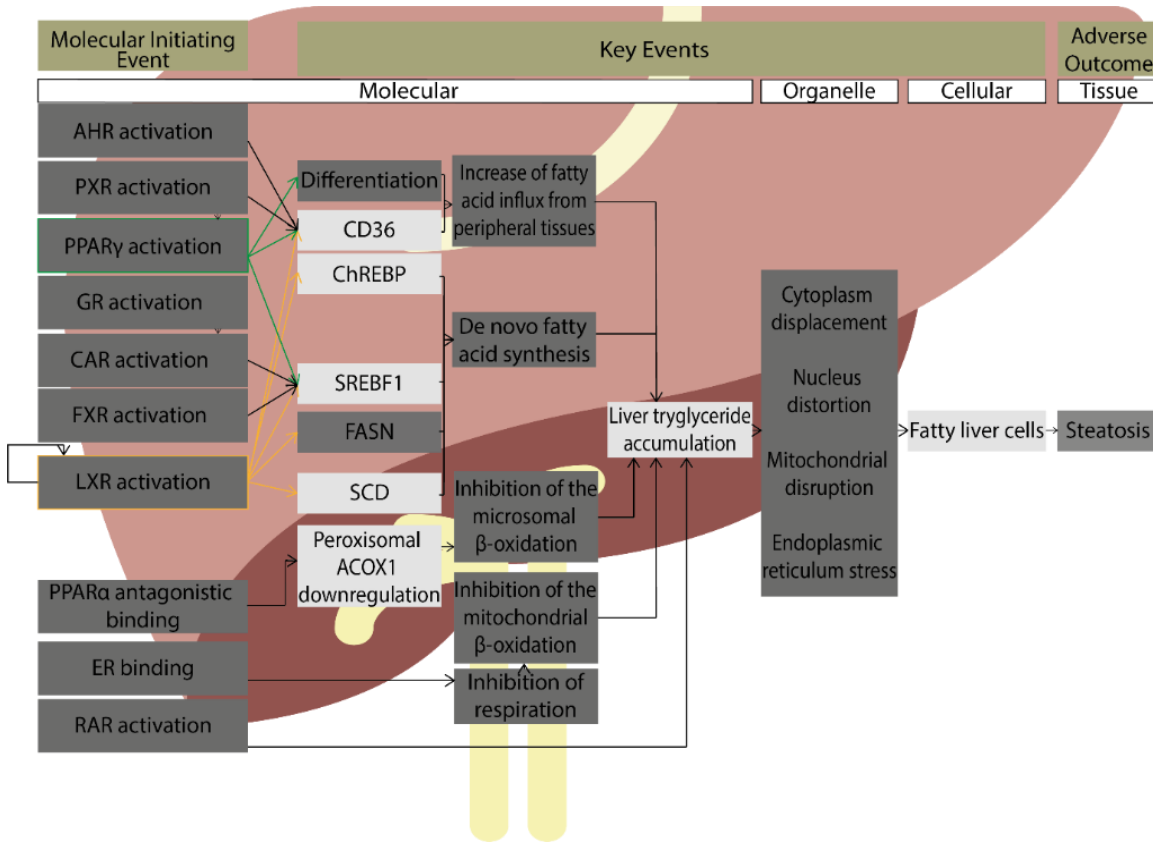
# Applying *In Vitro* Chemical Distribution Model

## Polycyclic aromatic hydrocarbons: phenanthrene

Serum	0%	2%	5%
Measured free	21%	8%	5%
Modeled free	32%	9%	5%
Measured in cells	10%	4%	2%
Modeled in cells	14%	5%	3%

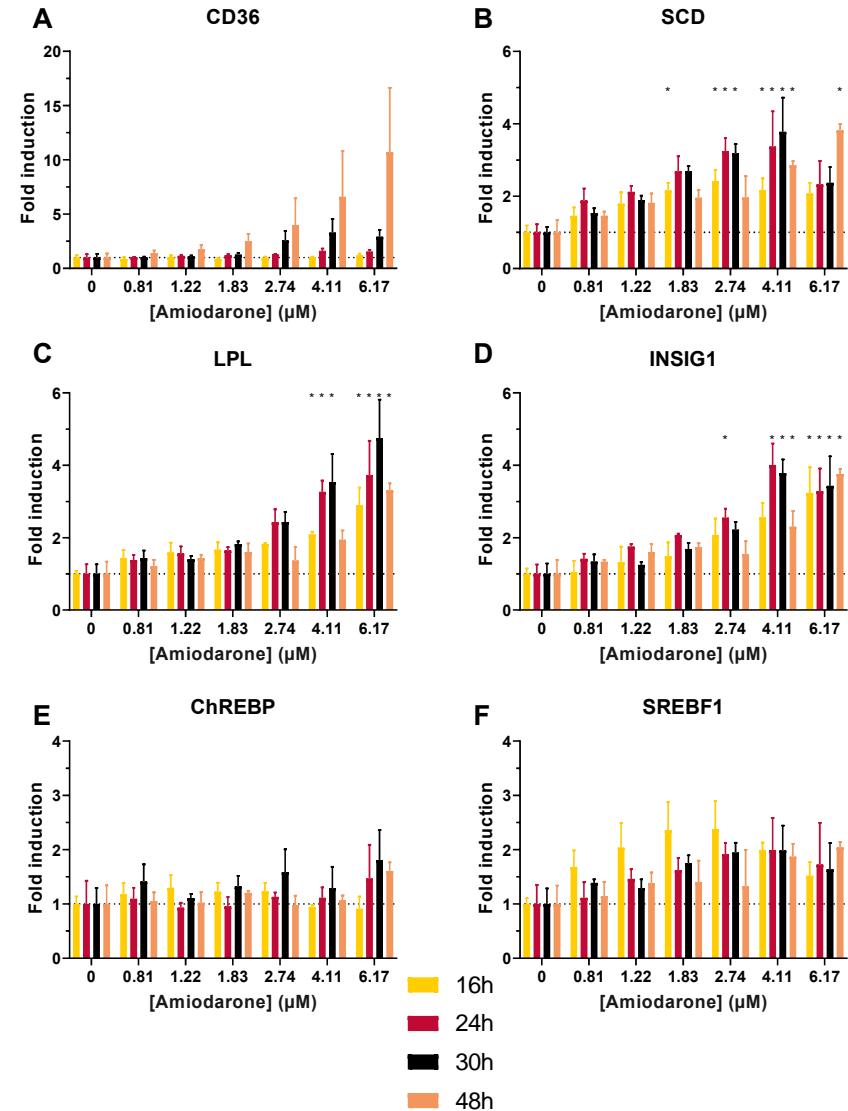


# *In vitro* test battery for KE perturbation



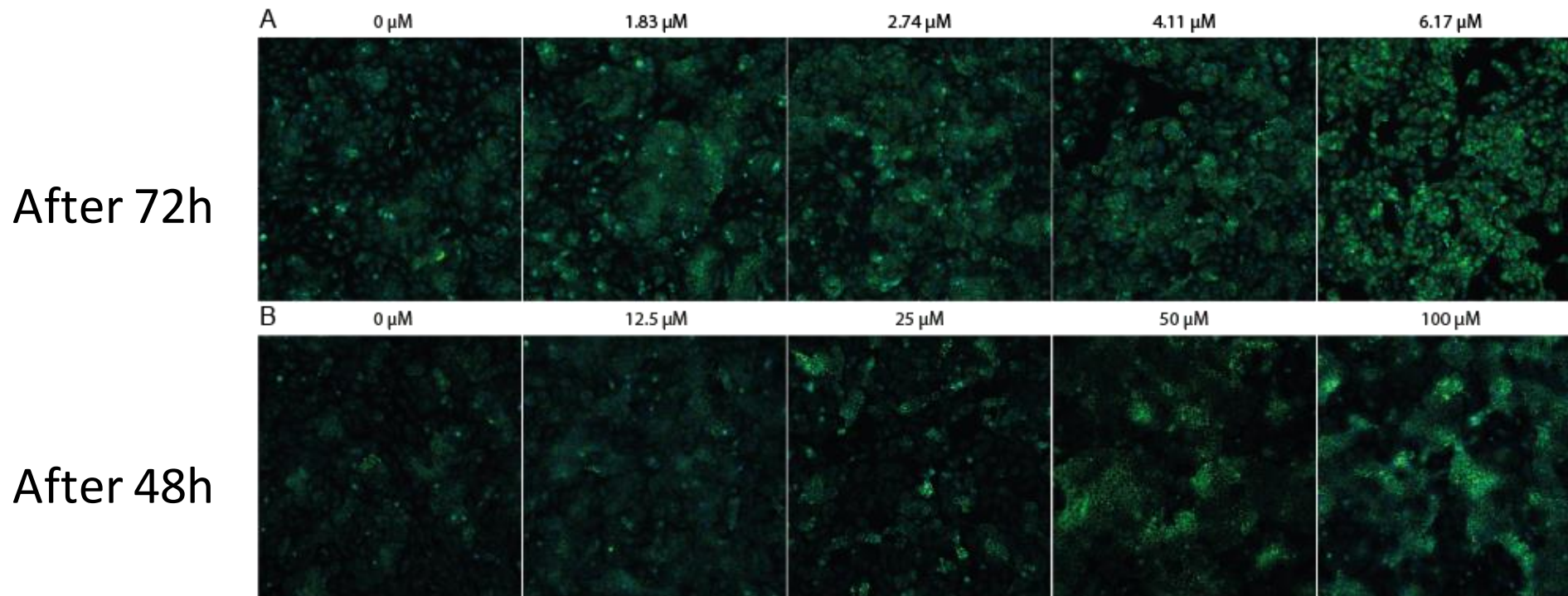
Kasteel et al., manuscript submitted

$T_{max}$  is KE dependent



# $T_{max}$ is chemical dependent...

## ■ Amiodarone (A) vs tetracycline (B)



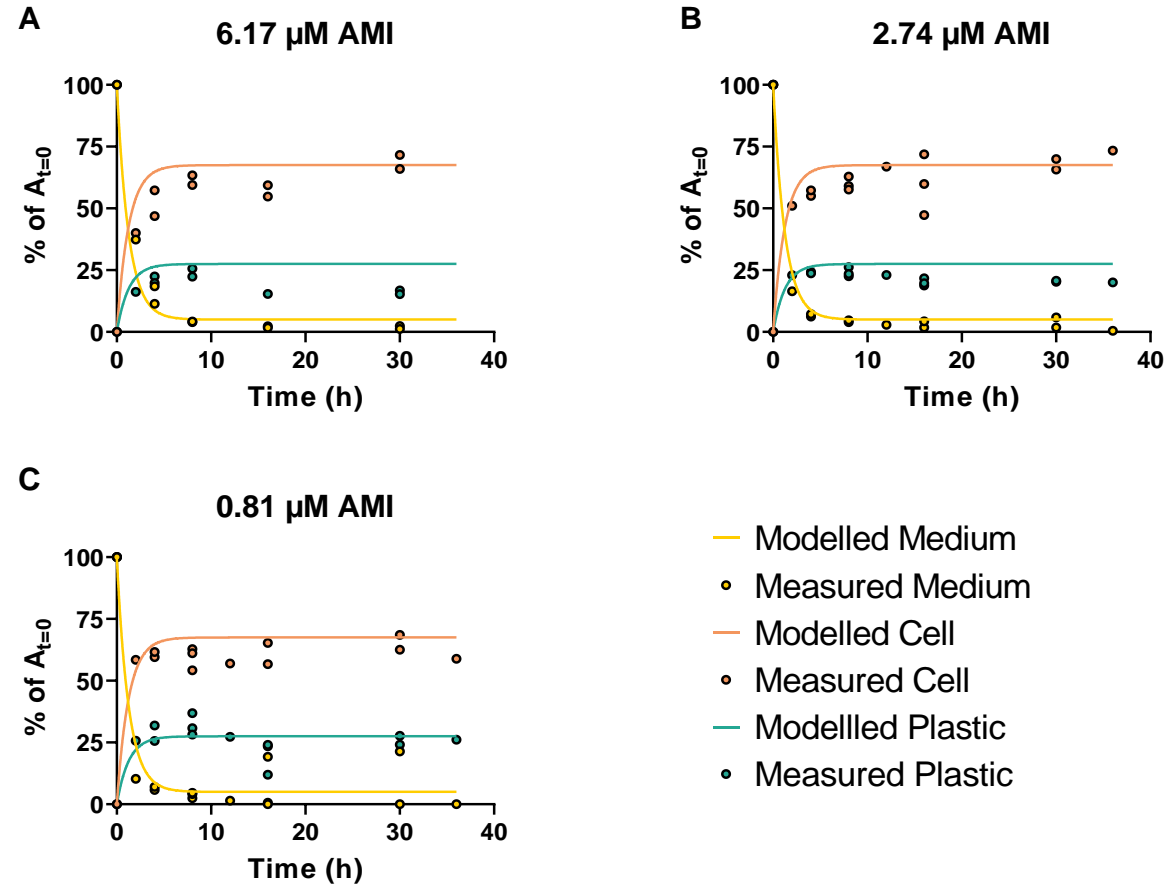
Sequence of KE may be misrepresented when using concentration response relationships of different chemicals



# *In vitro* kinetics needed for response-response modelling...

Use concentration-effect relationships ...

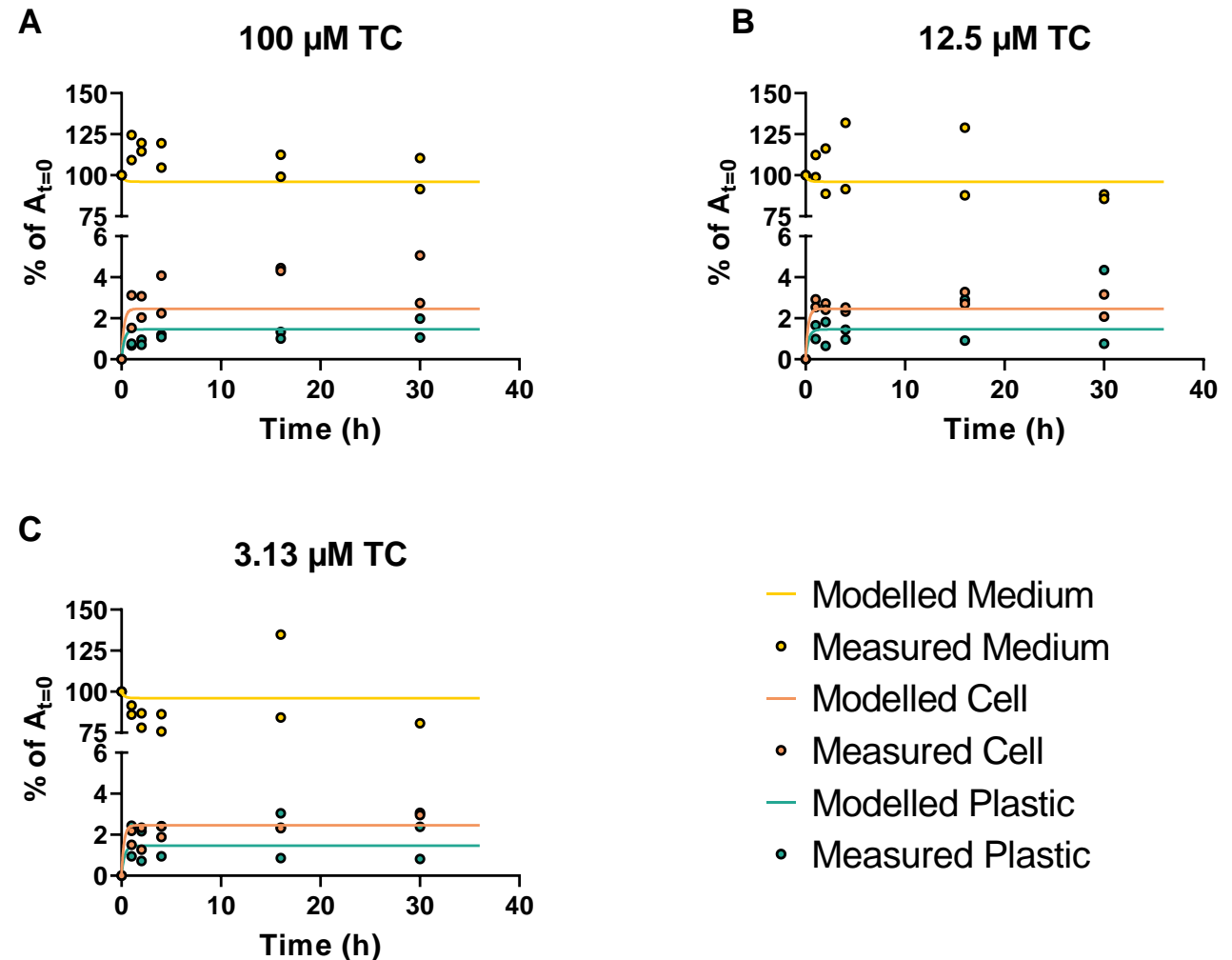
- at exposure time point leading to lowest EC
- based on internal cell concentrations



# *In vitro* kinetics needed for response-response modelling...

Use concentration-effect relationships ...

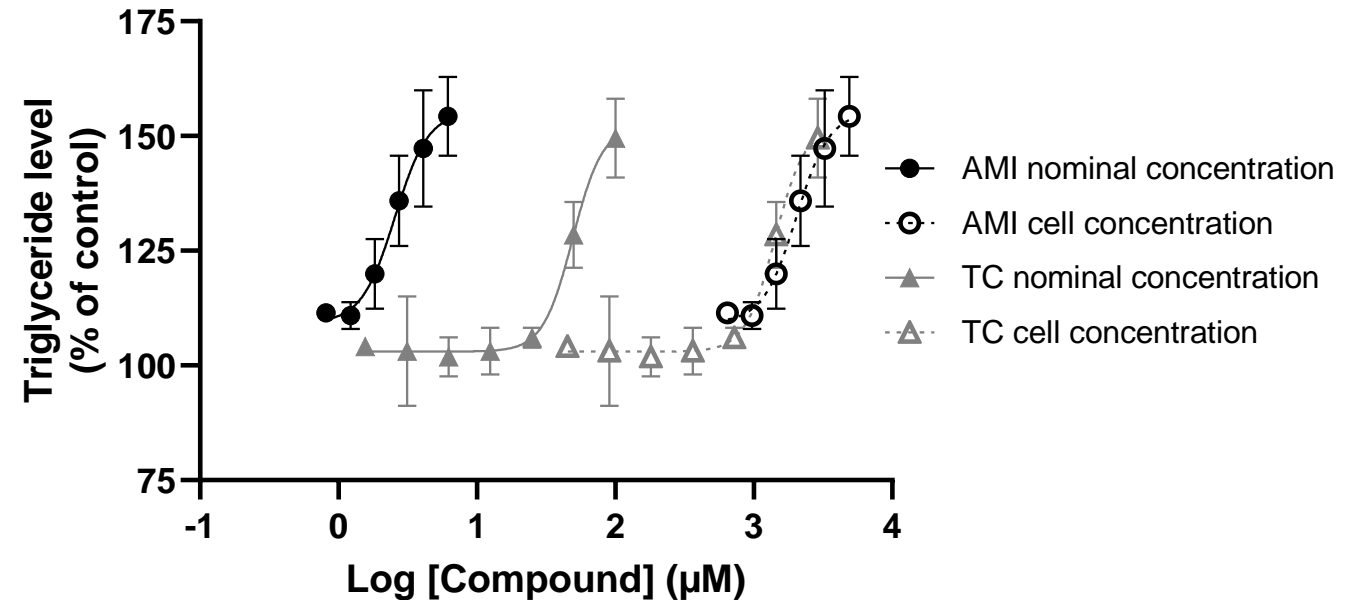
- at exposure time point leading to lowest EC
- based on internal cell concentrations



# *In vitro* kinetics needed for response-response modelling...

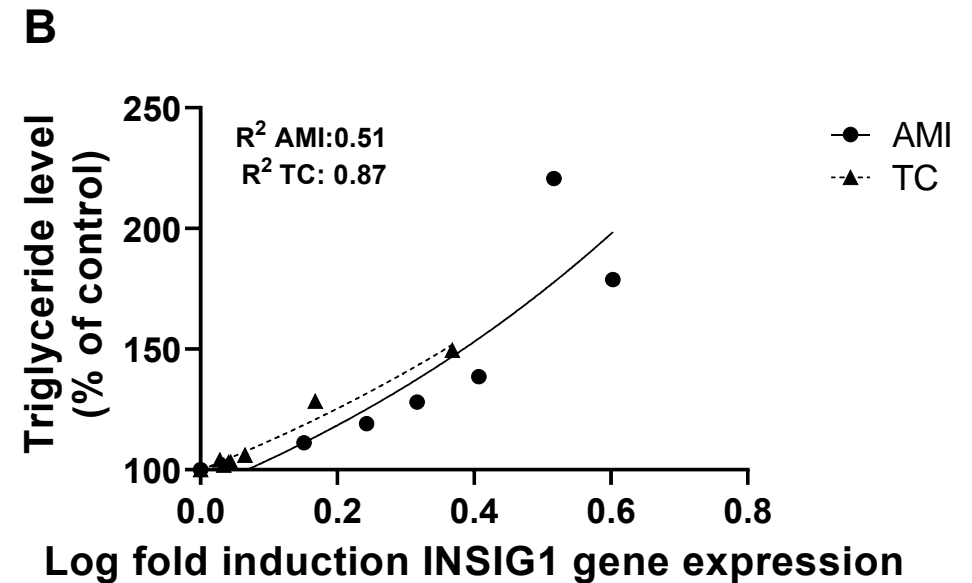
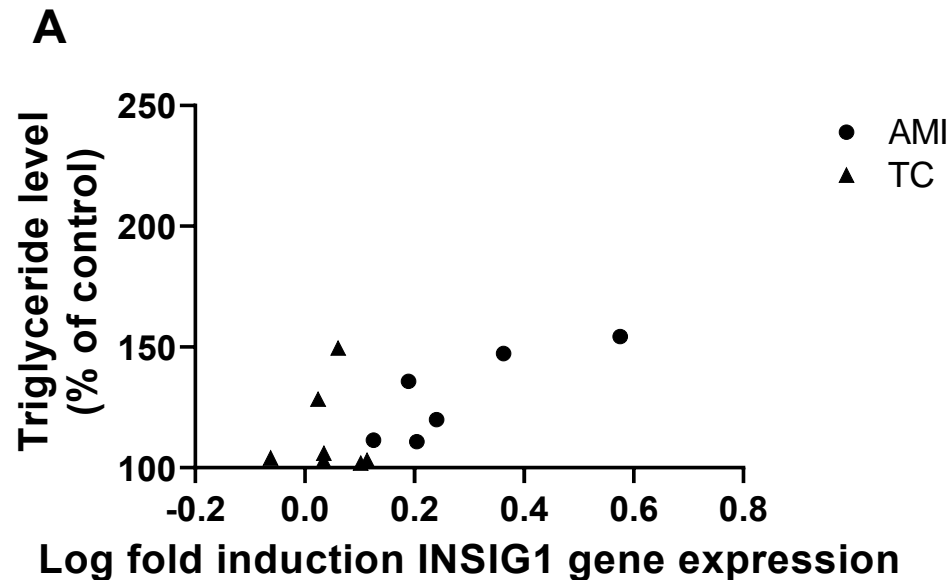
Use concentration-effect relationships ...

- at exposure time point leading to lowest EC
- based on internal cell concentrations



# *In vitro* kinetics needed for response-response modelling...

- Standard: readout @24h exposure vs @exposure at T<sub>max</sub> & using C<sub>cell</sub>

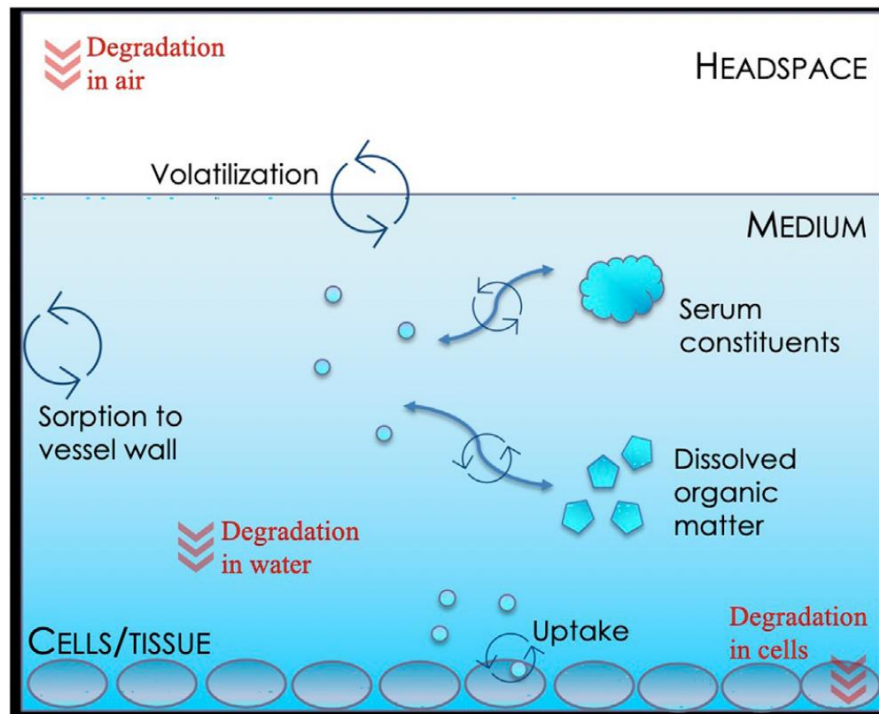


# Dynamic *In Vitro* Distribution Kinetics Modelling



frontiers | Frontiers in Toxicology

ORIGINAL RESEARCH  
published: 22 August 2022  
doi: 10.3389/ftox.2022.911128



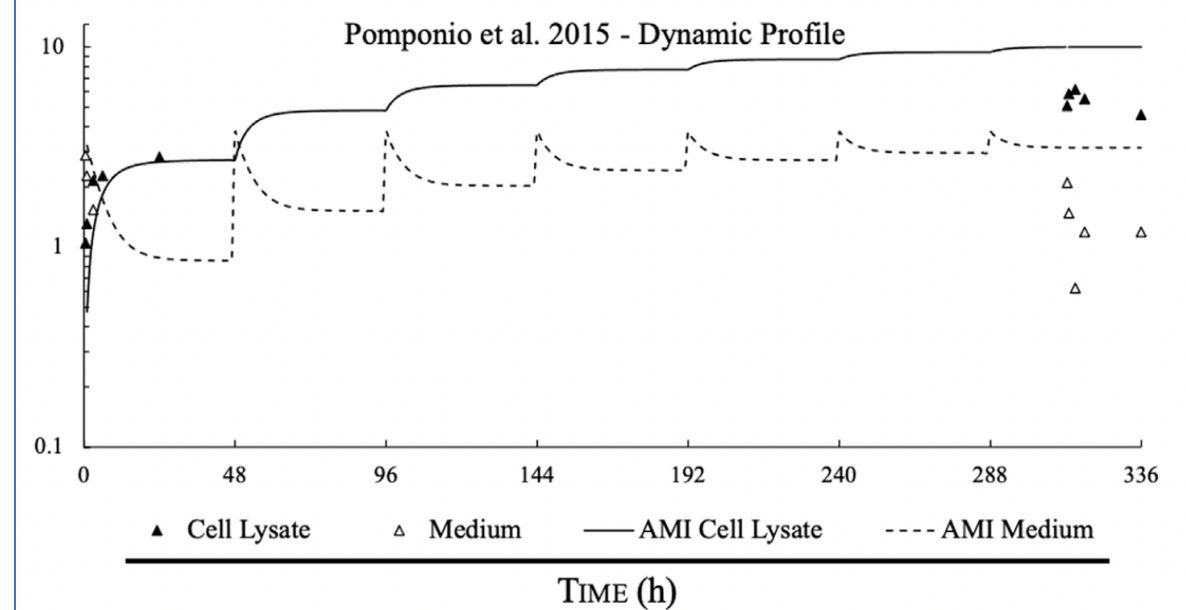
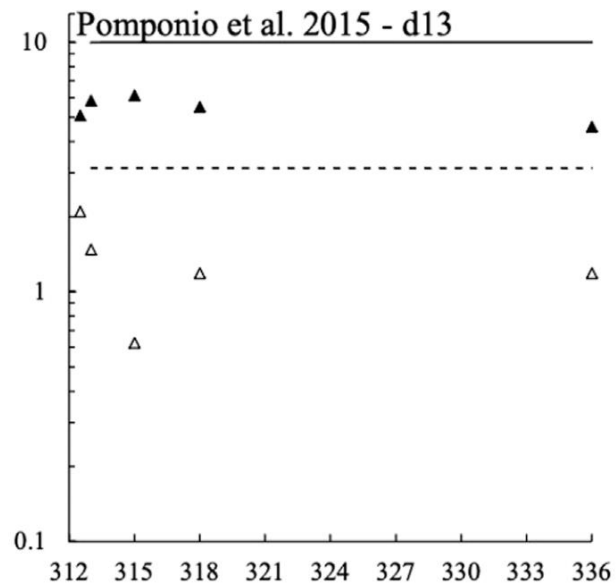
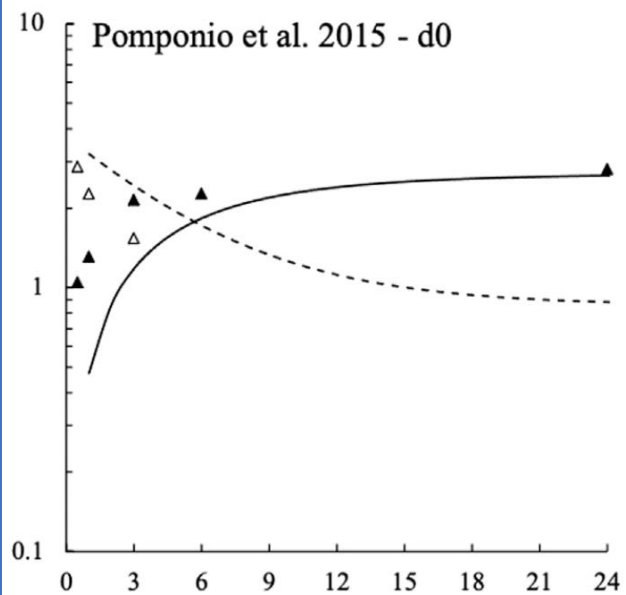
## Dynamic Mass Balance Modeling for Chemical Distribution Over Time in *In Vitro* Systems With Repeated Dosing

Sherri Bloch<sup>1,2</sup>, Jon A. Arnot<sup>3,4</sup>, Nynke I. Kramer<sup>5</sup>, James M. Armitage<sup>6</sup> and Marc-André Verner<sup>1,2\*</sup>

<sup>1</sup>Department of Occupational and Environmental Health, School of Public Health, Université de Montréal, Montreal, QC, Canada, <sup>2</sup>Centre de Recherche en Santé Publique, Université de Montréal et CIUSSS du Centre-Sud-de-l'Île-de-Montréal, Montreal, QC, Canada, <sup>3</sup>Department of Physical and Environmental Sciences, University of Toronto Scarborough, Scarborough, ON, Canada, <sup>4</sup>ARC Arnot Consulting and Research, Inc., Toronto, ON, Canada, <sup>5</sup>Division of Toxicology, Wageningen University, Wageningen, Netherlands, <sup>6</sup>AES Armitage Environmental Sciences, Inc., Ottawa, ON, Canada

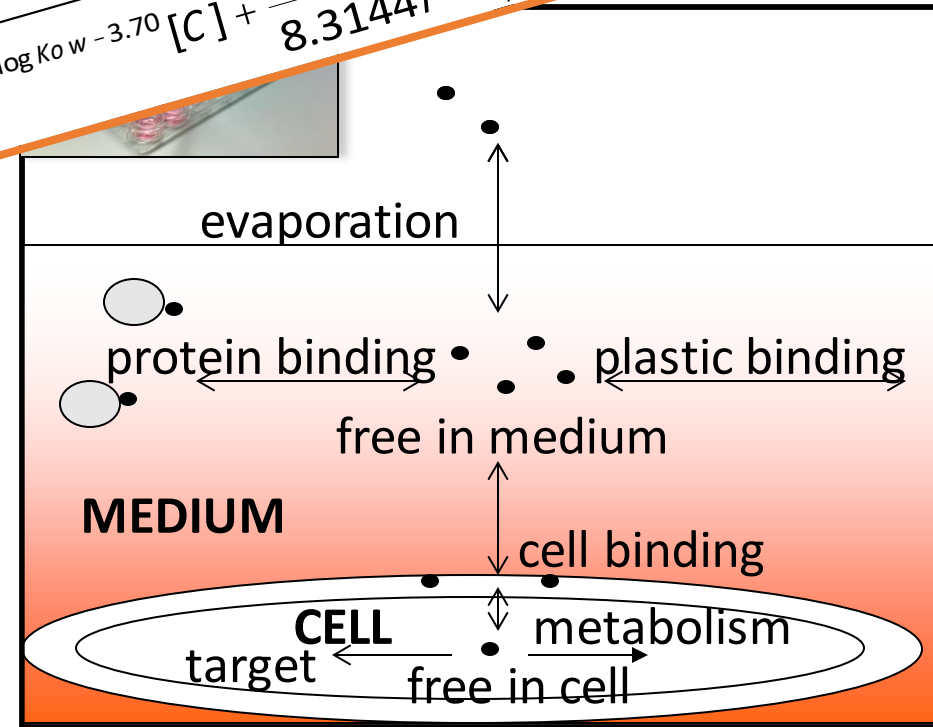
# Dynamic *In Vitro* Distribution Kinetics Modelling

Amiodarone (nmol/well)



# Modelling Evaporation

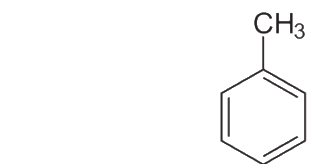
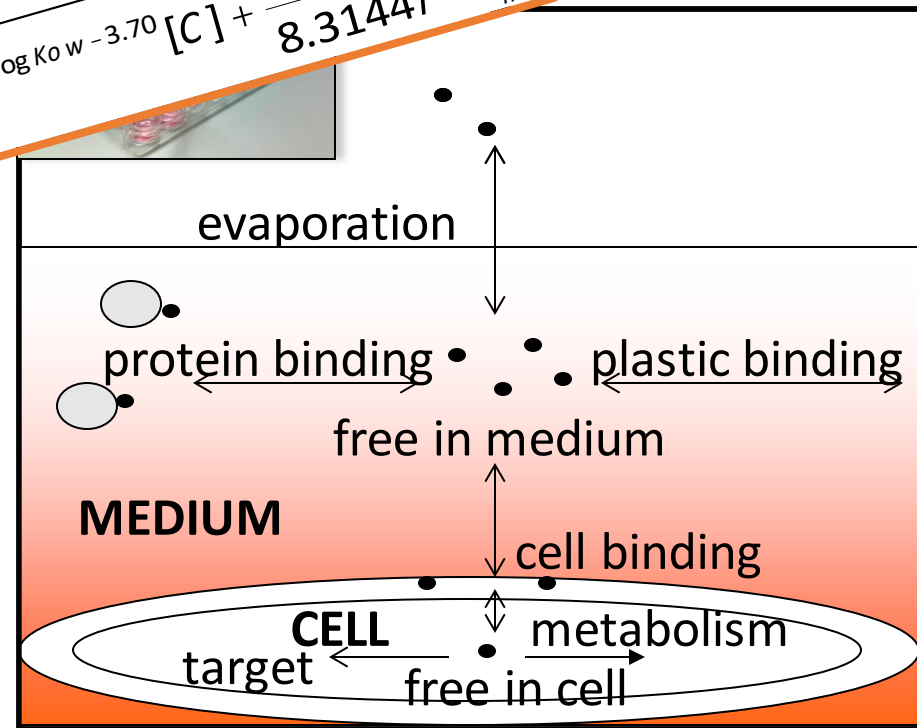
$$F = \frac{1}{1 + 10^{0.37 \log K_{ow} - 0.29} [S] + 10^{0.97 \log K_{ow} - 6.94} [P] + 10^{1.25 \log K_{ow} - 3.70} [C] + \frac{H}{8.3144T} \times \frac{V_a}{V_m}}$$



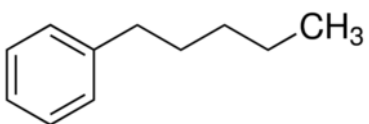
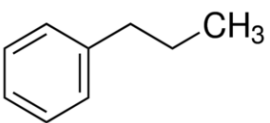


# Modelling Evaporation of Alkylbenzenes

$$F = \frac{1}{1 + 10^{0.37 \log K_{ow} - 0.29} [S] + 10^{0.97 \log K_{ow} - 6.94} [P] + 10^{1.25 \log K_{ow} - 3.70} [C] + \frac{H}{8.3144T} \times \frac{V_a}{V_m}}$$



Alkylbenzene	Modelled free
Toluene	43%
Propylbenzene	29%
Pentylbenzene	15%

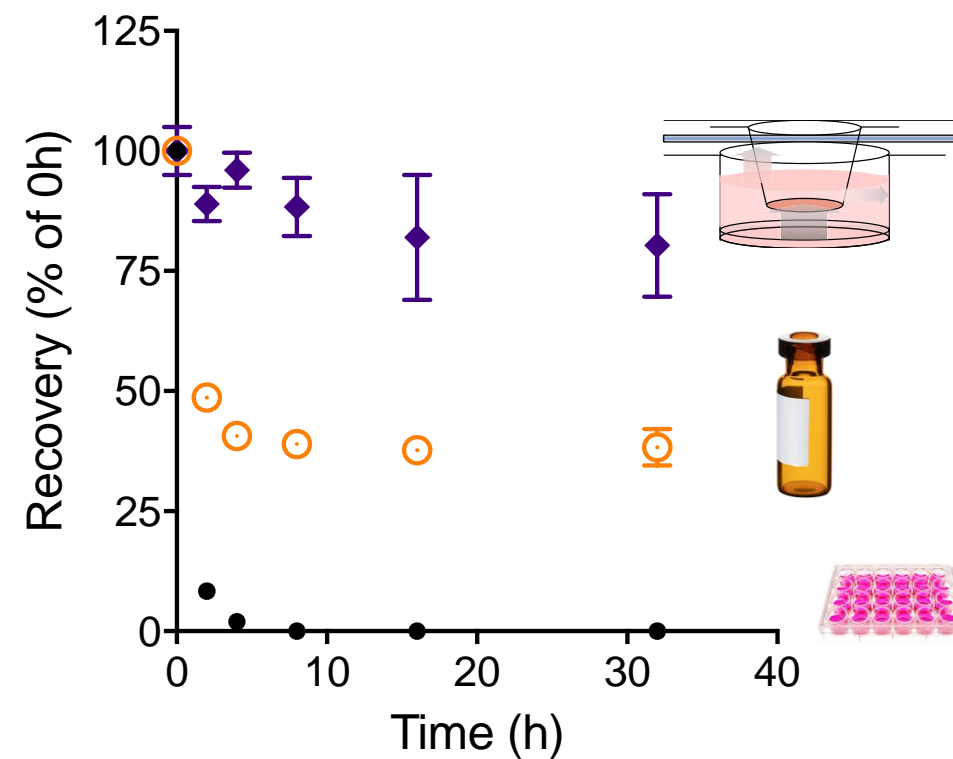


# Alternative Dosing Methods

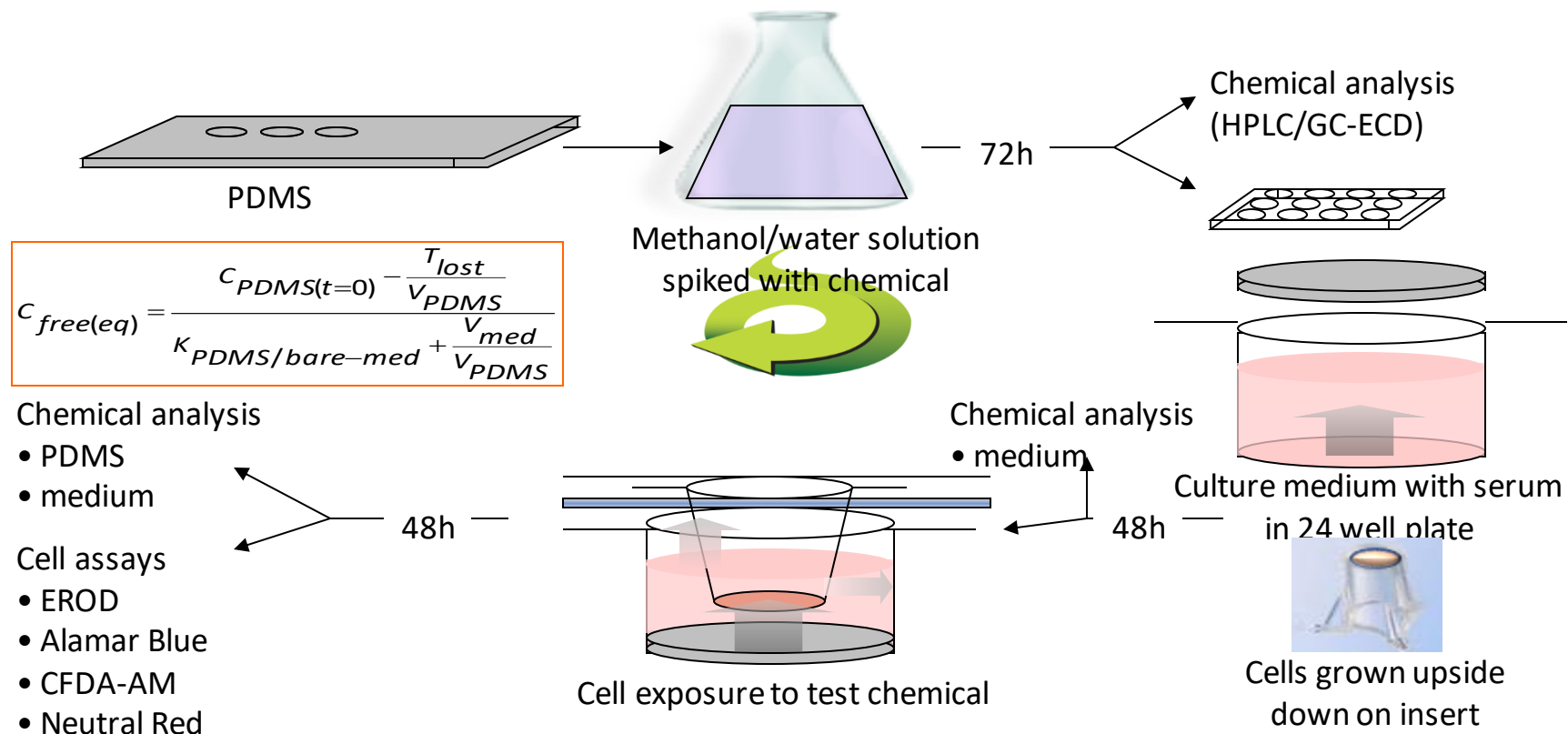
$$F = \frac{1}{1 + 10^{0.37 \log K_{ow} - 0.29} [S] + 10^{0.97 \log K_{ow} - 6.94} [P] + 10^{1.25 \log K_{ow} - 3.70} [C] + \frac{H}{8.3144T} \times \frac{V_a}{V_m}}$$



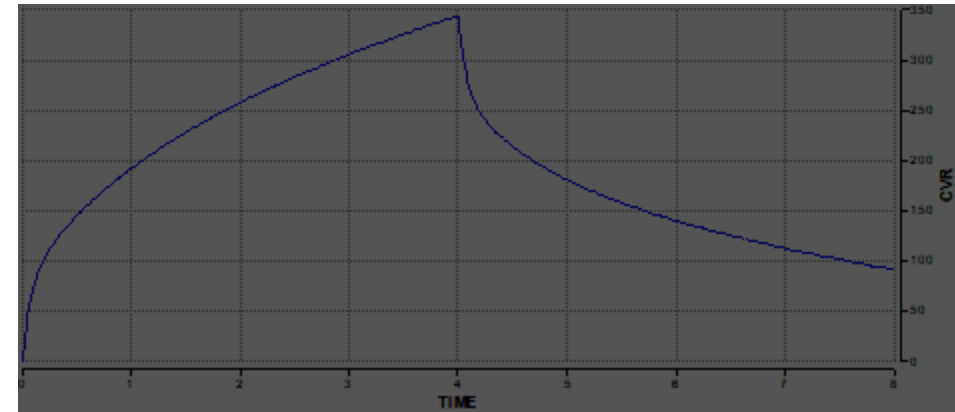
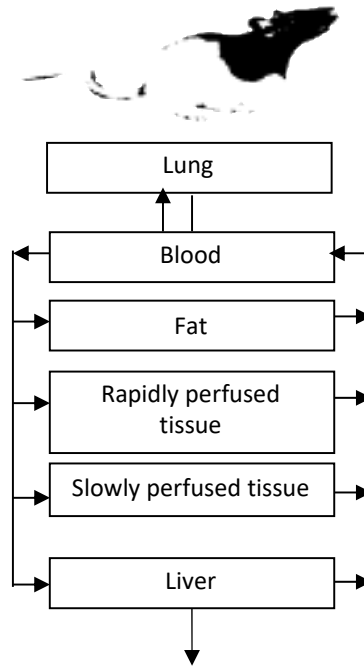
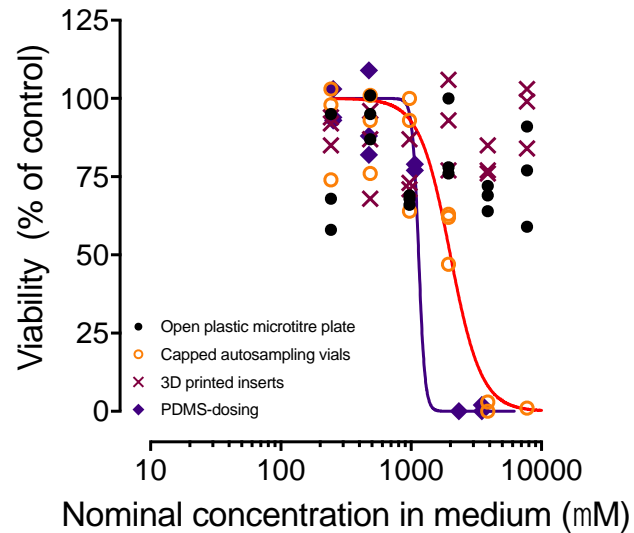
Alkylbenzene	Modelled free	Measured free	Measured free
Toluene	43%	0%	40±6%
Propylbenzene	29%	0%	27±3%
Pentylbenzene	15%	0%	17±1%



# Partition Controlled Dosing



# Alternative Dosing Methods and QIVIVE

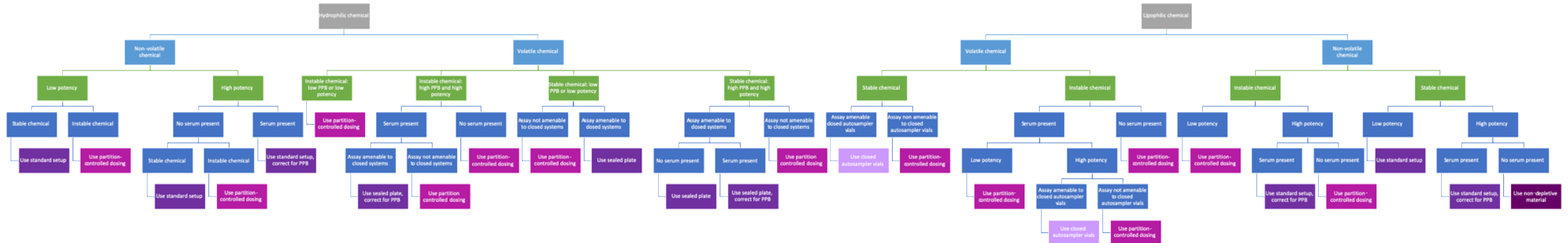


Est. 1230 ppm vs measured 1600 ppm  
LD50 1h inhalation in open chamber

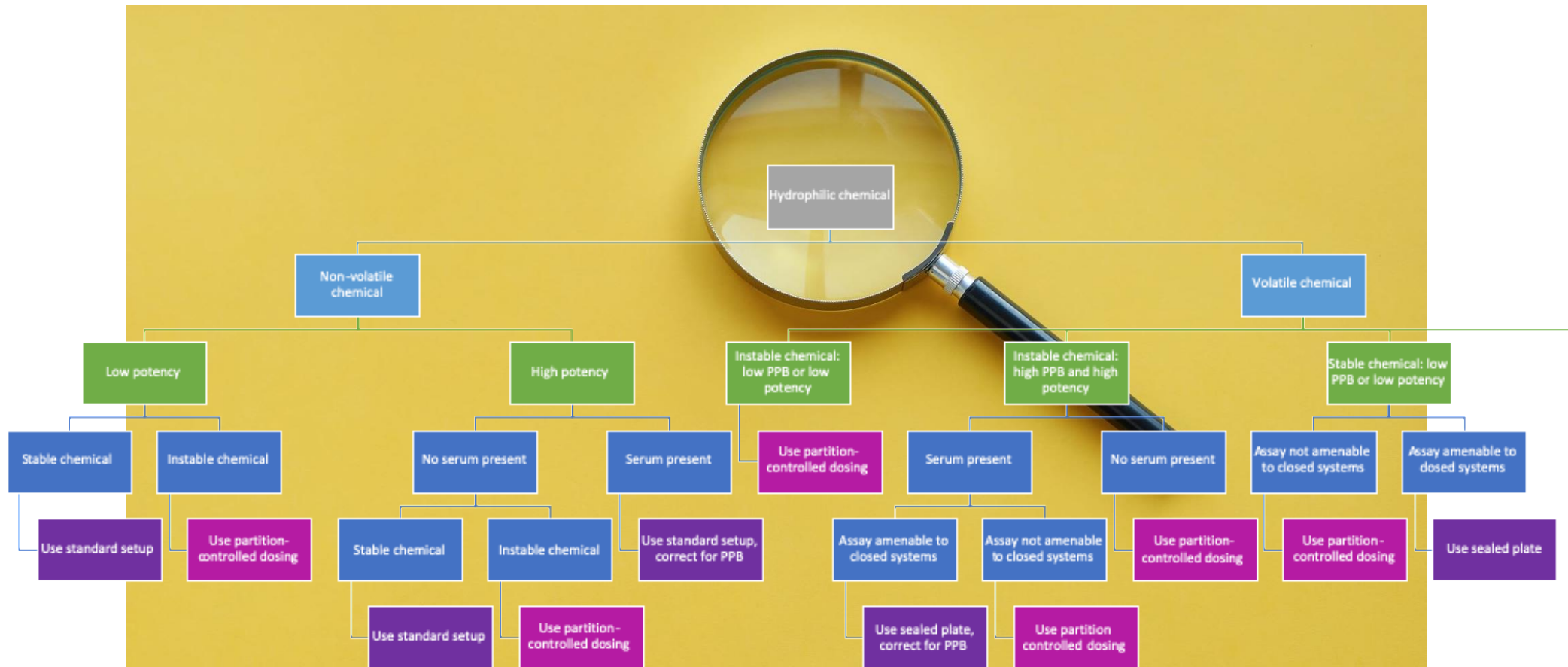
EC50 vials free (est.): 760  $\mu$ M  
LC50<sub>acute</sub> fish: 400  $\mu$ M

# Decision Tree

So when should you worry about nominal concentrations *in vitro*?



# When is Exposure 'Out of Control'?



# Conclusion

Choose your *in vitro* dose carefully!

